

STIC-ILL

RC261.H1 06

From: Seharaseyon, Jegatheesan
Sent: Wednesday, December 04, 2002 10:59 AM
To: STIC-ILL
Subject: 09/604,378

Adms

Importance: High

Please obtain the following articles. Thanks.

1. ACCESSION NUMBER: 2000159268 MEDLINE
DOCUMENT NUMBER: 20159268 PubMed ID: 10694945
TITLE: Prevention and treatment of chemotherapy- and
radiotherapy-induced oral mucositis: a review.
AUTHOR: Plevova P
CORPORATE SOURCE: Department of Radiotherapy, University Hospital of Ostrava,
Ostrava-Poruba, Czech Republic.. pavlina.plevova@fnspo.cz
SOURCE: ORAL ONCOLOGY, (1999 Sep) 35 (5) 453-70. Ref: 225
Journal code: 9709118. ISSN: 1368-8375.
PUB. COUNTRY: ENGLAND: United Kingdom
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)
General Review; (REVIEW)

2. ACCESSION NUMBER: 95211031 MEDLINE
DOCUMENT NUMBER: 95211031 PubMed ID: 7696971
TITLE: IL-11, a pleiotropic cytokine: exciting new effects of
IL-11 on gastrointestinal mucosal biology.
AUTHOR: Keith J C Jr; Albert L; Sonis S T; Pfeiffer C J; Schaub R G
CORPORATE SOURCE: Genetics Institute, Inc., Cambridge, Massachusetts.
SOURCE: STEM CELLS, (1994) 12 Suppl 1 79-89; discussion 89-90.
Journal code: 9304532. ISSN: 1066-5099.
PUB. COUNTRY: United States

3. ACCESSION NUMBER: 2000453978 MEDLINE
DOCUMENT NUMBER: 20464824 PubMed ID: 11012229
TITLE: The clinical development of recombinant human interleukin
11 (NEUMEGA rhIL-11 growth factor).
AUTHOR: Kaye J A
CORPORATE SOURCE: Clinical Research/Hematology, Genetics Institute, Inc.,
Cambridge, Massachusetts 02140, USA.
SOURCE: STEM CELLS, (1996) 14 Suppl 1 256-60. Ref: 26
Journal code: 9304532. ISSN: 1066-5099.
PUB. COUNTRY: United States
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)
General Review; (REVIEW)
(REVIEW, TUTORIAL)

4. DOCUMENT NUMBER: 97392673 PubMed ID: 9245489
TITLE: Mitigating effects of interleukin 11 on
consecutive courses of 5-fluorouracil-induced ulcerative
mucositis in hamsters.
AUTHOR: Sonis S T; Van Vugt A G; McDonald J; Dotoli E;
Schwertschlag U; Szklut P; Keith J
CORPORATE SOURCE: Division of Oral Medicine Oral and Maxillofacial Surgery,
and Dentistry, Brigham & Women's Hospital, Boston, MA
02115, USA.
SOURCE: CYTOKINE, (1997 Aug) 9 (8) 605-12.

Seyon.

J. Seharaseyon
Art Unit 1647
CM1 10D16
10B19 MB
Phone: (703)-305-1112
Fax: (703)-746-5177



Prevention and treatment of chemotherapy- and radiotherapy-induced oral mucositis: a review

P. Plevová*

Department of Radiotherapy, University Hospital of Ostrava, tř. 17 listopadu 1790, 708 52 Ostrava-Poruba, Czech Republic

Received 3 February 1999; accepted 23 March 1999

Abstract

Oral mucositis is a distressing toxic effect of systemic chemotherapy with many commonly utilized drugs and of head and neck irradiation in patients with cancer. The agents and methods that have been used and studied in chemotherapy- and radiotherapy-induced oral mucositis, their mechanisms of action, and the current knowledge of their efficiency to reduce the incidence, severity or shorten the duration of oral mucositis are reviewed in this article. Oral cooling is a cheap and available method to lower the severity of bolus 5-fluorouracil-induced oral mucositis. However, more effective methods are needed. Results of studies with granulocyte-macrophage colony-stimulating factor or granulocyte colony-stimulating factor are promising. Lasers are partly beneficial, but equipment-demanding. Modification of the chemotherapy regimen resulting in shortening of the exposition time to chemotherapy agents or chromomodulation of chemotherapy has been shown to lower mucosal toxicity of some regimens. Results of animal studies with locally applied transforming growth factor β 3 and interleukin-11 are also promising. Based on the findings of the role of the inflammatory cascade in the response of normal tissues to chemotherapy and radiotherapy, anti-inflammatory drugs might be beneficial. At the present time, no agent has been shown to be uniformly efficacious and can be accepted as standard therapy of chemotherapy- and radiotherapy-induced oral mucositis. Further intensive research is needed. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: Stomatitis prevention; Stomatitis control; Antineoplastic agents; Chemically induced; Radiotherapy-adverse effects

1. Introduction

Oral mucositis is a distressing toxic effect of systemic chemotherapy with many commonly utilized drugs and of head and neck irradiation in patients with cancer. Due to severe painfulness, oral mucositis interferes with the patient's quality of life and nutrition. It also increases the risk of systemic infections in immunocompromised patients due to disrupted barriers and often is the dose-limiting factor interfering with the intensification of anticancer therapy [1–3].

Oral mucositis is a consequence of the toxic effects of chemotherapeutic agents and irradiation on oral mucosa cells [2,4,5]. A complex hypothesis has been proposed as to the mechanism by which mucositis develops and resolves; it is based on four phases: an initial inflammatory/vascular phase; an epithelial phase; an ulcerative/bacteriological phase; and a healing phase [5]. The

inflammatory response induced in the involved tissues by chemotherapy and ionizing radiation occurs through the activation of intracellular and intercellular signaling pathways regulating gene expression of specific proteins involved in immune and inflammatory processes, such as cytokines, adhesion molecules, etc. [6–9]. The specific intracellular damage induced by cytotoxic drugs and responsible for the epithelial phase of mucositis development is generally well characterized [5]. In addition, recent studies have shown that most anticancer agents and γ irradiation kill cells by a common death program called apoptosis, which is the usual form of physiologic cell death in eukaryotic cells directed by a machinery that consists of molecular pathways discovered during the past several years [10–12]. Apoptosis may be involved in the development of chemotherapy- and radiotherapy-induced oral mucositis.

The role of oral bacterial colonization in the development of chemotherapy- and radiotherapy-induced oral mucositis is controversial. The appearance and resolution of oral mucositis is often considered to be linked to

* Tel.: +420-69-6984370 or +420-69-6717841; fax: +420-69-6919010.
E-mail address: pavlina.plevova@fnspo.cz

chemotherapy-induced neutropenia [13-15], which predisposes patients to oral infections, that could aggravate severity or prolong duration of oral mucositis [16]. The role of infection in the etiology of toxic oral mucositis is supported by the findings of abnormal oral flora colonization [17-22]. The role of gram-negative bacteria or endotoxin (lipopolysaccharide) may be in potentiation of the immune response of the tissues to chemotherapeutic agents or irradiation, since endotoxin is a potent inflammation mediator [8,20]. On the other hand, however, oral mucositis complicates chemotherapy regimens that are relatively nonmyelotoxic; its grade or recovery does not correlate with the presence, severity or recovery of neutropenia in some reports [16,23-27]; and locally applied antimicrobials, desinfectants and antimycotics failed to substantially influence our ability to prevent or heal it [28-33].

The clinical appearance of oral mucositis may range from mild discomfort and erythema to painful erythema, and oedema and/or ulcerations [2,21].

The agents and methods that have been used and studied in chemotherapy- and radiotherapy-induced oral mucositis, their mechanisms of action, and the current knowledge of their ability to reduce the incidence, severity or shorten the duration of oral mucositis are reviewed in this article (Table 1). As many drugs have been studied and tried with either intention, prevention and treatment are dealt with together. Symptomatic therapy of this complication is beyond the scope of this paper and is reviewed elsewhere [2,3,34].

2. Locally applied measures and pharmacotherapeutics

Dental restoration to healthy status, including therapy of dental caries, periodontal disease, detection of foci, and correction of any other potential sources of irritation, such as ill-fitting prostheses and orthodontic appliances, is advised before or early in cancer therapy in order to reduce the frequency of oral problems [35-39]. Although a retrospective study has shown a substantial decrease in the frequency of oral mucositis in the period of aggressive dental interventions compared to the previous one, when no such measures were used [36], results of a prospective study focused on the superiority of an intensive dental care and oral hygiene protocol were not clinically impressive [40].

Oral hygiene programs are commonly advised to reduce the amount and activity of oral microflora and to prevent or reduce discomfort associated with oral mucositis [2,20,35,41-44]. Patients are instructed about an effective and frequent mechanical plaque removal by daily brushing with a soft toothbrush, flossing with dental floss and rinsing with saline solution and mild solution of sodium bicarbonate, about lip lubrication, 'sugarfree' products to enhance oral moistness or

saliva substitutes, when needed [2,35,37,38,41]. An early beginning of proper oral care reduced oral toxicity of radiotherapy in a pilot study [45]. Although statistically significant, the superiority of an intensive oral hygiene protocol involving initial treatment of dental lesions and tooth and gum brushing during aplasia was not clinically useful compared to a limited oral hygiene protocol in bone marrow transplantation (BMT) patients [40].

Sucralfate is a basic aluminum salt of sucrose octasulfate useful in the treatment of peptic ulcer disease [46]. The mechanism of action of this drug is not known with certainty, but it may involve the binding of sucralfate to the damaged mucosal surface proteins and the formation of a protective coating over ulcers with possible epithelial regeneration and angiogenesis-promoting properties [47-50]. Anecdotal reports suggested pronounced responses to oral sucralfate in patients with chemotherapy-induced oral mucositis [51-53], supported by results of several studies [54-58]. However, there was seen no statistically significant reduction in mucositis in other, randomized double-blind studies [59-63]. Sucralfate can reduce the experience of pain [55,59,60,62,63]. Another mouth-coating agent, *kaolin pectin*, combined with diphenhydramine, which is a H1-histamine antagonist and local anesthetic, was able to reduce oral pain without reducing the degree of mucositis [64].

Vitamin E probably acts as an antioxidant; it presumably inhibits oxidation of essential cellular constituents and prevents the formation of toxic oxidation products [65-67]. Reactive oxygen intermediates are important in the activation of inflammatory tissue response [9]. Although the antioxidant effects of vitamin E are relatively weak [68], duration of chemotherapy-induced oral mucositis was significantly shorter in patients treated topically with vitamin E than in the placebo group in two small studies [69,70].

Tretinoin (all-trans-retinoic acid), a vitamin A derivative, is known to stimulate wound healing [71]; it has anti-inflammatory effects that may create better local circumstances for healing [72]; besides that, it induces epithelial growth via cell differentiation modulation [73]. It has been reported to significantly reduce the severity of oral mucositis in BMT patients [74].

Hydrogen peroxide, once recommended as an oral rinse to aid in the management of adhesive mucus and for its antimicrobial properties [75,76], came into disrepute because of its antifibroblast healing-delaying and possible carcinogenic activity [18,41]. In addition, frequent mechanical cleansing of the mouth has been reported more beneficial than hydrogen peroxide [44].

Chlorhexidine is a powerful broadly active antimicrobial and antiseptic. Assuming that oral mucositis in patients undergoing intensive antineoplastic treatments results largely from superinfection of a reversibly

Table 1

Drugs and methods studied in the prevention and/or therapy of chemotherapy- or radiotherapy-induced oral mucositis (survey of published studies)^a

Reference No.	No. of patients (study/control group)	Cause	Randomized/controlled/double-blind?	P/T	Study characteristics	Result—impact on oral mucositis
I. Locally applied measures and pharmacotherapeutics						
a. Dental restoration						
[36]	495	CHT	No/no/no	P	Dental intervention in standardized protocols; compared to a published study without these measures	Substantial reduction in frequency
[40]	150 (75/75)	CHT (ev. BMT)	Yes/yes/no	P	Intensive versus limited protocol (i.e. with vs without initial treatment of dental lesions, tooth and gum brushing)	Significant reduction, not clinically impressive
b. Oral hygiene						
[45]	30 (10/10/10)	RT	Yes/yes/no	P	Instructions on oral care given at 1 day versus 1 week before RT versus no instructions	Significant reduction in the 2nd group
[40]	150 (75/75)	CHT (ev. BMT)	Yes/yes/no	P	Intensive versus limited protocol (i.e. with vs without initial treatment of dental lesions, tooth and gum brushing)	Significant reduction, not clinically impressive
c. Mouth-coating agents:						
•Sucralfate						
[51]	4	CHT	No/no	T		Accelerated healing
[52]	23	CHT	No/no	T, P		Reduction
[53]	18	CHT	No/no	T		Reduction mostly
[54]	46 (23/23)	5-FU-CHT	Yes/yes/yes	P	Sucralfate versus placebo	Significant reduction
[55]	45 (24/21)	RT	Yes/yes/no	P	Sucralfate yes versus no	Significant reduction
[56]	40 (20+20)	RT	No/no	P, T	P: sucralfate, later + fluconazole, T: sucralfate + fluconazole	Reduction
[57]	48 (24/24)	RT	Yes/yes/yes	P	Sucralfate versus placebo	Significant temporary reduction
[58]	45	5-FU	No/no	P		Low incidence
[59]	48 (24/24)	CHT	Yes/yes/yes	P + T	Sucralfate versus placebo	No difference (less pain)
[60]	33 (16/17)	RT	Yes/yes/yes	P + T	Sucralfate versus placebo	No difference (less pain)
[61]	40 (20/20)	RT	Yes/yes/yes	P	Sucralfate versus placebo	No difference
[62]	50 (27/23)	5-FU	Yes/yes/yes	T	Sucralfate versus placebo	No difference (less pain)
[63]	111 (53/58)	RT	Yes/yes/yes	T	Antacid + diphenhydramine + viscous lidocaine with versus without sucralfate	No difference (less pain)
•Kaolin-pectin						
[64]	29 (7/7/15)	RT	Yes/yes/yes	P + T	Oral hygiene + sucralfate versus + diphenhydramine + kaolin-pectin versus historical group without oral hygiene or rinse	No difference between study groups (less pain than in historical group)
d. Vitamins:						
•Vitamin E						
[69]	18 (9/9)	CHT	Yes/yes/yes	T	Vitamin E versus placebo	Significant reduction of duration
[70]	19 (10/9)	CHT (ev. BMT)	Yes/yes/no	P	Vitamin E versus placebo	Significant reduction in leukemia patients, no difference in other groups

(Table continued on next page)

Table 1 (continued)

Reference No.	No. of patients (study/control group)	Cause	Randomized/controlled/double-blind?	P/T	Study characteristics	Result—impact on oral mucositis
•Tretinoin [74]	11 (6/5)	CHT, CHRT (BMT)	Yes/yes/no	P	Tretinoin yes or no	Significant reduction of severity
<i>e. Antibiotics, desinfectantia:</i>						
•Hydrogen peroxide [44]	40 (20/20)	RT	Yes/yes/no	P	Hydrogen peroxide versus saline solution	Significantly worse outcome
•Chlorhexidin [77]	16 (8/8)	CHT	Yes/yes/yes	P	Chlorhexidine versus placebo	Significant reduction
[78]	51 (24/27)	CHRT (BMT)	Yes/yes/yes	P	Chlorhexidine versus placebo	Significant reduction
[79]	40 (19/21)	CHT	Yes/yes/yes	P + T	Chlorhexidine versus placebo	Significant reduction
[79]	30 (16/14)	RT	Yes/yes/yes	P + T	Chlorhexidine versus placebo	No difference
[80]	13 (6/7)	CHT (ev. BMT)	Yes/yes/yes	P	Chlorhexidine versus placebo	Significant reduction in BMT patients, no difference in non-BMT patients
[28]	100 (50/50)	CHRT (BMT)	Yes/yes/yes	P	Chlorhexidine versus placebo	No difference
[29]	30 (15/15)	RT	Yes/yes/yes	P + T	Chlorhexidine versus placebo	No difference
[30]	86 (34/16/18/18)	CHT, BMT	Yes/yes/no	P	Chlorhexidine + nystatin versus nystatin versus chlorhexidine versus saline solution	No difference
[31]	52 (25/27)	RT	Yes/yes/yes	P	Chlorhexidine versus placebo	Slight aggravation (more discomfort)
[81]	28 (14/14)	CHT	Yes/yes/no	P	Chlorhexidine yes versus no	No difference
[82]	222 (111/111)	CHT	Yes/yes/yes	P	Chlorhexidine versus placebo	No difference
•Povidone-iodine solution [83]	40 (20/20)	CHRT	Yes/yes/no	P	Povidone-iodine versus placebo	Significant reduction
•Selective decontamination [20]	15	RT	No/yes/no	P	Lozenges of polymyxine E, tobramycin, amphotericin B compared to historical controls (chlorhexidine, placebo) [39]	Significant reduction
[89]	59 (22/37)	RT	Yes/yes/no	P	Sucralfate + (ciprofloxacin or ampicillin) + clotrimazole versus sucralfate	Reduction
[22]	221 (112/109)	RT	Yes/yes/yes	P	Lozenges of polymyxin, tobramycin, amphotericin B versus placebo	No difference in severe grades incidence; significant distribution reduction
[32]	26 (12/14)	CHT, CHRT (BMT)	Yes/yes/no	T	Polymyxin E, tobramycin and amphotericin B with chlorhexidine versus diphenhydramine, magnesium-alumina, lidocaine	Significant reduction, not clinically impressive
[33]	112 (54/58)	RT	Yes/yes/yes	P + T	Lozenges of polymyxine E, tobramycin, amphotericin B versus placebo	No objective difference, significant reduction in patient-reported mucositis
<i>f. Anti-inflammatory agents:</i>						
•Chamomila [41]	20	RT	No/yes/no	P	Compared with a historical group without chamomila	Reduction compared to previous
	32	CHT	No/yes/no	T	Compared with a historical group without chamomila	Short duration
	46	CHT	No/yes/no	P	Compared with a historical group without chamomila	Low incidence
[91]	164 (82/82)	5-FU	Yes/yes/yes	P	Chamomila versus placebo	No difference
•Betamethasone [94]	5	RT	No/no	P	High-dose betamethasone	Total prevention of mucositis

Table 1 (continued)

Reference No.	No. of patients (study/control group)	Cause	Randomized/controlled/double-blind?	P/T	Study characteristics	Result—impact on oral mucositis
•Benzylamin						
[98]	67 (37/30)	RT	Yes/yes/yes	P, T	Benzylamine versus placebo	Significant reduction (less pain)
[99]	36 (19/17)	CHT, RT	Yes/yes/yes	P, T	Benzylamine versus placebo	Significant reduction
[100]	43 (25/18)	RT	Yes/yes/yes	P, T	Benzylamine versus placebo	Significant reduction
[101]	25 (13/12)	RT	Yes/no/no	P	Benzylamine versus chlorhexidine	No difference (more discomfort)
•Mesalazine						
[109]	14 (14 cc/14cc)	CHT (ev. BMT)	No/yes/no	T	Mesalazine yes versus no	(Less pain)
g. Cytokines:						
•GM-CSF						
[27]	24	CHT	No/no	T	GM-CSF at four various concentrations versus placebo	Reduction
[110]	45 (9/9/9/9/9)	CHT	Yes/yes/yes	P		No significant reduction, aggravation in low-dose drug groups; no dose response
•TGF-β3						
No studies in humans available						
•EGF						
[116]	12 (6/6)	CHT	Yes/yes/yes	T	EGF versus placebo	No difference
h. Eicosanoids:						
•PGE₁ (misoprostol)						
[118]	15 (8/7)	CHT (PSCT)	Yes/yes/yes	P	PGE ₁ versus placebo	Significant aggravation
•PGE₂						
[120]	8	CHT	No/no	P, T	PGE ₂ Yes or no	Successful prophylaxis and reduction
[121]	24 (10/14)	CHRT	No/yes/no	P		Significant reduction (less pain)
[122]	15	CHRT	No/no	T		No severe grades
[123]	60 (31/29)	CHT, CHRT (BMT)	Yes/yes/yes	P		No difference
i. Multiagent topical mouthrinses						
[18]	79	CHRT	No/no	P, T	Hydrogen peroxide + polyvinylpyrrolidon-iod + nystatin + 5% dexpanthenol solution Hydrocortison + nystatin + tetracycline + diphenhydramine versus placebo	Reduction, accelerated healing
[126]	12 (7/5)	RT	Yes/yes/yes	P		Significant reduction
j. Epitelization promoting agents:						
•Silver nitrate						
[128]	16 (16ss/16ss)	RT	No/yes/no	P	Applied to one side of buccal mucosa, the contralateral one served as control Applied to one side of buccal mucosa, the contralateral one served as control	Significant reduction
[130]	10 (10ss/10ss)	RT	No/yes/no	P		No difference
k. Antineoplastic agent antagonists:						
•Leucovorin						
[132]	14 (14cc/14cc)	HD-MTX	No/yes/no	P	Leucovorin + hyaluronidase yes versus no	No difference
[133]	13 (44cc)	HD-MTX	No/no	P		No benefit

(Table continued on next page)

Table 1 (continued)

Reference No.	No. of patients (study/control group)	Cause	Randomized/controlled/double-blind?	P/T	Study characteristics	Result—impact on oral mucositis
<i>1. Allopurinol</i>						
[139]	6 (6cc/6cc)	5-FU	No/yes/no	P	Allopurinol yes versus no	Substantial reduction
[140]	16	5-FU	No/no	P		Substantial reduction
[141]	44 (22/22)	5-FU	Yes/yes/yes	T	Allopurinol versus placebo	Significant reduction
[142]	77 (38/39)	5-FU	Yes/yes/yes	P	Allopurinol versus placebo	No difference
<i>2. Locally applied nonpharmacological methods</i>						
<i>a. Radiation shields</i>						
[144]	125 (61/64)	RT	Yes/yes/no	P	Midline mucosa-sparing blocks yes versus no	Significant reduction
<i>b. Oral cooling (cryotherapy)</i>						
[145]	95 (50/45)	5-FU	Yes/yes/no	P	Cryotherapy yes versus no	Significant reduction
[146]	84 (44/40)	5-FU	Yes/yes/no	P	Cryotherapy yes versus no	Significant reduction
[147]	20 (9/11)	5-FU-CHT	Yes/yes/no	P	Ice bar with fibrinolysin and deoxyribonuclease yes versus no	Significant reduction
[148]	178 (89/89)	5-FU	Yes/yes/no	P	Cryotherapy duration of 60 versus 30 min	No difference
[149]	22	L-PAM, CHT (BMT, PSCT)	No/no	P		Reduction in single-agent L-PAM, not in multiagent regimens
[150]	18	L-PAM	No/no	P		Reduction in both single agent and multiagent regimens
<i>c. Soft lasers</i>						
[153]	67 (25/21/21)	5-FU-CHT	No/yes/no	P, T	Laser prophylaxis/therapy yes versus no	Substantial reduction (prophylaxis), reduced duration (therapy)
[154]	59 (23/16/20)	5-FU-CHT	No/yes/no	P, T	Laser prophylaxis or therapy yes versus no	Reduction (prophylaxis group), accelerated healing (therapy group)
[155]	22 (22ss/22ss)	CHT, CHRT (BMT)	Yes/yes/yes	P	Laser on one buccal side versus sham treatment to the contralateral side	Significant reduction
[156]	30 (15/15)	CHRT (PSCT, BMT)	Yes/yes/yes	P	Laser versus sham treatment	Significant reduction
<i>3. Systemically applied pharmacotherapeutics</i>						
<i>a. Antioxidants:</i>						
•Beta carotene						
[157]	20 (10/10)	CHRT	Yes/yes/no	P	Beta-carotene enriched versus standard diet	Significant reduction
•Azelastrine						
[160]	63 (37/26)	5-FU-CHRT	Yes/yes/no	P	Vitamins C + E + glutathione with versus without azelastrine	Significant reduction, not clinically impressive
<i>b. Immunomodulatory drugs:</i>						
•Pentoxifylline						
[165]	50 (30/20)	CHT (BMT)	No/yes/no	P	Oral PTX versus historical controls without PTX	Significant reduction
[166]	92 (31/61)	CHT, CHRT (BMT)	No/yes/no	P	iv PTX versus historical controls without PTX	Significant aggravation
[167]	88 (44/44)	CHT, CHRT (BMT)	Yes/yes/no	P	Oral PTX versus placebo	No difference
[168]	140 (70/70)	CHT, CHRT (BMT)	Yes/yes/yes	P	Oral PTX versus placebo	No difference
[169]	49 (28/21)	CHT, CHRT (BMT)	No/yes/no	P	Oral PTX versus historical controls without PTX	No difference

Table 1 (continued)

Reference No.	No. of patients (study/control group)	Cause	Randomized/controlled/double-blind?	P/T	Study characteristics	Result—impact on oral mucositis
[170]	10 (10cc/10cc)	5-FU-CHT	Yes/yes/yes	P	Oral PTX versus placebo	No difference
Indomethacin						
[171]	19 (10/9)	RT	Yes/yes/yes	P	Indomethacin versus placebo	Significant delay of onset
Immunoglobulin						
[172]	124 (79/45)	RT, CHRT	No/yes/no	T	IMIG yes versus no	Reduction
[173]	86	RT	No/no	T	IMIG	Shortened duration
[174]	81 (39/42)	RT	Yes/yes/yes	T	IMIG versus placebo	Significant reduction
[175]	42 (22/20)	RT, CHRT	No/yes/no	P	IMIG yes versus no	Significant reduction in CHRT patients, no difference in RT patients
[176]	3 (3cc/9 cc)	HD-MTX	No/yes/no	P	IVIG yes versus no	No mucositis in IVIG cc
<i>c. Anticholinergic drugs:</i>						
Propantheline						
[180]	12 (6/6)	Eto-CHT (BMT)	Yes/yes/no	P	Propantheline versus placebo	Significant reduction
[181]	31	Eto-CHT (PSCT)	No/no	P	Control—published reference without propantheline	Reduction
Atropine						
[132]	6 (6cc/6cc)	HD-MTX	No/yes/no	P	Atropine yes versus no	No benefit
<i>d. Cytokines:</i>						
G-CSF, GM-CSF						
[23]	22 (18/6)	CHT	No/yes/no	P	G-CSF yes versus no	Significant reduction
[182]	33 (15/18)	CHT (BMT)	No/yes/no	P	G-CSF yes versus no	Significant reduction of duration
[183]	22 (22cc/22cc)	CHT (ev. BMT)	No/yes/no	P	GM-CSF yes versus no	Reduction
[184]	37 (23/14)	CHT	Yes/yes/no	P	GM-CSF versus placebo	Significant reduction
[185]	37	5-FU	No/no	P	GM-CSF	Reduction
[186]	26 (10/16)	CHT (BMT)	No/yes/no	P	G-CSF yes versus no	No difference
[187]	80 (41/39)	CHT	Yes/yes/no	P	G-CSF yes versus no	No difference
[188]	50 (20/30)	CHT, CHRT (BMT)	No/yes/no	P	G-CSF yes versus no	No difference
[16]	20 (9/11)	5-FU-CHT	Yes/yes/no	P	GM-CSF yes versus no	Significant reduction
[25]	26 (14/12)	CHT, CHRT (BMT)	No/yes/no	P	G-CSF yes versus no	Significant duration reduction (TBI-patients)
[189]	109 (53/56)	CHRT (BMT)	Yes/yes/yes	P	GM-CSF versus placebo	Significant reduction
[190]	14 (7/7)	CHT	Yes/yes/no	P	G-CSF yes versus no	Significant reduction
[191]	10	RT	No/no	P	GM-CSF	Low incidence
[192]	29	CHT, RT	No/no	P	GM-CSF	Reduction
<i>Interleukin-11</i>						
No studies in humans available						
<i>e. Antiviral drugs:</i>						
Acyclovir						
[201]	60 (28/23/9)	CHT, CHRT (BMT)	No/no	P	Acyclovir in HSV seropositive, seronegative and unknown serology patients	No difference
[203]	15 (12/3)	BMT	No/no	P	Acyclovir in seropositive patients	Majority of patients developed mucositis
[204]	34 (16/18)	CHT, RT	Yes/yes/yes	P	Acyclovir versus placebo	No difference
[205]	90 (45/45)	CHT	Yes/yes/yes	P	Acyclovir versus placebo	No difference
<i>f. Metabolic substrate supplementation:</i>						
Glutamine						
[212]	14 (14cc/14cc)	CHT	No/yes/no	P	Oral glutamine yes versus no	Significant reduction
[213]	13 (13cc/13cc)	CHT	Yes/yes/yes	P	Oral glutamine versus placebo	Significant reduction
[214]	20cc (10/10)	CHT	Yes/yes/yes	P	Total parenteral nutrition with versus without glutamine	No difference
[215]	28 (14/14)	5-FU	Yes/yes/yes	P	Oral glutamine versus placebo	No difference

Table 1 (continued)

Reference No.	No. of patients (study/control group)	Cause	Randomized/controlled/double-blind?	P/T	Study characteristics	Result—impact on oral mucositis
g. Hormone:						
•Melatonin						
[216]	80 (40/40)	CHT	Yes/yes/no	P	Melatonin yes versus no	No difference
4. Other methods:						
a. Modification of the chemotherapy regimen						
[217]	43 (21/22)	HD-MTX	Yes/yes/no	P	4-h infusion of 12 g/m ² versus 36-h infusion of 1 g/m ² 24-h infusion weekly, compared to published studies with 120- or 96-h infusion forth or third weekly, respectively	Significant reduction in the 1st group Substantial reduction
[26]	42	5-FU-CHT	No/no	P		
b. Chronotherapy						
[221]	186 (93/93)	5-FU-CHT	Yes/yes/no	P	Chronotherapy versus constant rate infusion	Significant reduction

^a P, prophylactic use; T, therapeutic use; CHT, chemotherapeutic agents other than 5-fluorouracil, methotrexate, melphalan, etoposide; ev. BMT, BMT performed in some patients; RT, radiotherapy; 5-FU, 5-fluorouracil; 5-FU-CHT, 5-fluorouracil-based chemotherapy; CHRT, chemotherapy; GM-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor; TGF- β , transforming growth factor β ; EGF, epidermal growth factor; PGE_{1,2}, prostaglandin E₁, E₂; CHT (BMT, PSCT) or CHRT (BMT, PSCT), bone marrow or peripheral stem cell transplantation conditioning regimen; amphi B, amphotericine B; ss, buccal sides; cc, cycles; HD-MTX, high-dose methotrexate; L-PAM, melphalan; PTX, pentoxifylline; iv, intravenous; IMIG, intramuscular immunoglobulin; IVIG, intravenous immunoglobulin; Eto, etoposide; Eto-CHT, etoposide-based chemotherapy; TBI, total body irradiation; HSV, herpes simplex virus.

damaged mucosal barrier, the efficiency of chlorhexidine to reduce this complication has been intensively studied. It has been shown to be effective in several studies [77-80]; in other studies, however, there was no clinically demonstrable benefit of chlorhexidine in the prevention and treatment of irradiation- or chemotherapy-induced or BMT-related oral mucositis [28-31,79,81,82], although improved oral hygiene and some changes in oral mucosa colonization have been observed in chlorhexidine users [28-30,78]. Mouthwash-induced discomfort has been reported in the chlorhexidine arm in some studies [31,81].

Povidone-iodine solution, a disinfection agent, significantly decreased radiochemotherapy-induced oral mucositis in a study [83].

The results of various studies suggest that yeast colonization is not involved in the pathogenesis of mucositis [84-86] and identical observations were reported for viridans streptococci [29,85]. As a high oral carriage of gram-negative bacilli was found in several studies [22,29,87,88], a hypothesis was established based on the role of pathological gram-negative oral flora, maybe particularly of endotoxin, in the etiology of radiotherapy- and chemotherapy-induced oral mucositis and the concept of *selective decontamination* has been developed [20,21]. In a pilot study, lozenges containing polymyxin E, tobramycin, and amphotericin B have been used in irradiated head and neck cancer patients; successful elimination of oral gram-negative bacilli has

been achieved and severe forms of irradiation oral mucositis have been prevented when compared retrospectively with patients using placebo or chlorhexidine [20]. Addition of ciprofloxacin or ampicillin and clotrimazole to the sucralfate mouthwash also reduced radiation mucositis [89]. In other studies, selective decontamination did not reach clinically significant reduction in mucositis [22,32,33].

Chamomila, a solution prepared from the flower of the chamomile plant, contains mainly chamazulene, levomenol, polyins, and flavonoids. The combination of these constituents has anti-inflammatory and spasmolytic effects and promotes granulation and epithelization [90]. Carl and Emrich [41] have observed that Kamillolan Liquidum mouthwashes applied prophylactically delayed onset and reduced severity of radiation mucositis, and prevented the occurrence of severe mucositis in the majority of the patients who received systemic chemotherapy when compared with historical controls. The results of a phase III trial did not show any benefit of chamomila in 5-fluorouracil-induced oral toxicity, however [91].

Glucocorticosteroids are anti-inflammatory drugs inhibiting synthesis of inflammatory proteins with the involvement of complex mechanisms at the molecular level, which leads to the decrease in transcription of genes involved in inflammation [9,92,93]. The mucosa of five patients, who used a high-dose betamethasone mouthwash before and during radiotherapy

for malignant parotid tumors, showed progressive whitening as radiation progressed, which was in contrast to the erythema that usually occurs. The mucosa remained virtually ulcer-free and no discomfort or bleeding of oral mucosa appeared. Histological examination of the oral mucosa taken at completion of irradiation showed absence of the inflammatory component of irradiation-induced mucositis and some thickening and keratinization of the buccal epithelium [94].

Benzydamine is a non-steroidal drug that possesses topical analgesic, anaesthetic, anti-inflammatory, and antimicrobial properties [95-97]. Reduction in mucositis has been reached using benzydamine [98-100]; on the other hand, there was no reduction in another study [101]. Significant relief of pain has been observed in benzydamine users [98,102,103]. However, oral burning associated with benzydamine rinses has been noted frequently [101,103,104].

Mesalazine (5-aminosalicylic acid) is an anti-inflammatory agent [105]; salicylates have been shown to inhibit the activation of one of the transcription factors involved in the transcription of inflammatory genes, albeit only in relatively high concentrations [106]. Topical mesalazine is used in the therapy of inflammatory bowel disease [105] and has been shown to be effective in the treatment of oral aphthous ulcers [107] and of oral and genital ulceration in Behcet's disease [108]. Results of a preliminary study of topical mesalazine in the treatment of chemotherapy- and radiotherapy-induced oral mucositis showed some benefit associated with the application of this agent [109].

Studies with locally applied granulocyte-macrophage colony-stimulating factor (GM-CSF) gave conflicting results so far [27,110]. GM-CSF may have a direct stimulatory effect on the growth or regeneration of oral mucosa cells [16].

The beta transforming growth factors (TGFs- β) transiently inhibit the cycling of epithelial cells in G1 phase [111]. The TGF- β 3 administration reduces proliferation of oral epithelium in vitro and in vivo and may thus decrease the susceptibility of these cells to the cytotoxic effect of chemotherapy. Topical application of TGF- β 3 to oral mucosa in a hamster model prior to chemotherapy significantly reduced the incidence, severity and duration of oral mucositis [112,113].

Epidermal growth factor (EGF) has a variety of effects on epithelial proliferation, differentiation, and chemotaxis [114]. Its level has been shown to decrease in oral secretions during radiation therapy and correlate inversely with the degree of oral mucositis [115]. EGF mouthwash was not observed to accelerate healing of chemotherapy-induced oral mucositis in a small group of patients, however [116]. Moreover, hamsters receiving EGF had significantly more severe and prolonged mucositis. Despite a hypothesis that EGF exposure later may hasten re-epithelization, delayed exposure to EGF

only appeared to delay the onset of mucositis and had no beneficial effects on either the severity or duration of mucositis [34].

Misoprostol is a synthetic prostaglandin E_1 analog with greater mucosal-protective activity than natural prostaglandins. It is used to prevent and treat gastrointestinal lesions induced by nonsteroidal anti-inflammatory drugs and upper gastrointestinal ulcerations [117]. In another study, misoprostol exhibited adverse effects [118].

Prostaglandin E_2 (PGE_2) is believed to possess cytoprotective properties [119]. Preliminary results suggested that topical application of PGE_2 (dinoprost) could be effective in the prevention of anticancer therapy-related oral mucositis [120-122]. In a double-blind prospective clinical trial, however, there was no significant difference in mucositis between the BMT patients dissolving PGE_2 in the mouth and those treated with placebo. In patients receiving PGE_2 , a significantly higher incidence of herpes simplex virus reactivation has been documented. In the patients with herpes simplex virus infection receiving PGE_2 , the incidence of severe mucosal lesions was significantly higher than in patients without herpes simplex virus infection who had received PGE_2 or in the control group [123]. Although PGE_2 is considered to possess anti-inflammatory properties [124], there are reports suggesting that endogenous PGE_2 has an important regulatory role in oral inflammation [125]. Exogenous PGE_2 may also have pro-inflammatory effects, responsible for the observed adverse effects associated with herpes simplex virus infection.

Various multiagent topical mouthrinses are used in the management of radiotherapy- or chemotherapy-induced oral mucositis. These include antimicrobial and antifungal agents combined with epithelization-promoting or mouth-coating agents, or anti-inflammatory drugs, or local anaesthetics; for instance, a combination of hydrogen peroxide plus betaisodone plus nystatine plus dexpantenol [18], or hydrocortison plus nystatine plus tetracycline plus diphenhydramine [126], or magnesium aluminum hydroxide plus diphenhydramine plus viscous lidocaine, or nystatin plus lidocaine plus solucortef plus sucralfate plus syrup alta [34]. However, the efficacy of these regimens is difficult to assess and long-term studies supporting such treatment are not available [127].

Maciejewski et al. [128,129] have reported that oral mucosa burning with 2% solution of silver nitrate a few days before radiotherapy reduced the severity of mucositis. They suggested that silver nitrate can stimulate the unirradiated mucosa into a more effective proliferative state before the start of radiotherapy. In another study, local conditioning of human oral mucosa by 3% silver nitrate solution significantly increased epithelial proliferation rate in healthy volunteers. Despite this fact, the results concerning oral mucositis were not reproduced [130].

Leucovorin (5-formyltetrahydrofolate) is used to protect normal tissues from the toxic effect of high-dose methotrexate, a folic acid antagonist [131]. Leucovorin mouthwash was expected to protect oral mucosa cells by antagonizing locally the inhibition of purin- and thymidilatesynthesis induced by methotrexate. However, in several studies, topical application of leucovorin did not effectively prevent development of oral mucositis [132,133]. The saliva concentrations of methotrexate and 7-hydroxymethotrexate have been shown not to correlate with the development of oral mucositis [134].

Allopurinol is a structural isomer of hypoxanthine. Certain findings suggested that oxypurinol, the major metabolite of allopurinol, could attenuate 5-fluorouracil (5-FU)-induced toxicity by inhibiting specific enzymes involved in the formation of toxic 5-FU metabolites [135-138]. Based on these findings, an assumption was made that allopurinol mouthwash might attenuate the toxicity of 5-FU on oral mucosa. Clark and Slevin [139] have observed a decrease in 5-FU-induced oral mucositis in six patients when allopurinol mouthwash was used and some studies [140,141] supported their findings. However, a randomized, placebo-controlled, double-blind study was closed preliminary, because a planned interim analysis showed convincingly negative results [142].

3. Locally applied nonpharmacological methods

Shields can be constructed to protect uninvolved oral tissues during radiation [3,143,144]. They have been shown to significantly reduce acute toxicity of radiotherapy in the oral region [144].

Oral cooling (cryotherapy) probably causes local vasoconstriction and, thus, temporarily reduces oral mucosal blood flow and the amount of the drug delivered to oral mucosa cells. Patients swish ice chips in their mouths or mouthwash with ice-cold water for a total of 30 min, starting 5 min prior to each dose of the drug. Cryotherapy has been shown to significantly reduce bolus 5-FU-induced oral mucositis [145-147]. A 60-min duration of cryotherapy does not provide more benefit than a 30 min one [148]. 5-FU is a drug with short plasma half-life; this method cannot be used for patients treated by continuous 5-FU infusion, however [16]. Protective effect of cryotherapy procedure as mentioned or in the form of ice-pop eating has been also observed in patients receiving high-dose melphalan. Besides vasoconstriction, a perhaps temperature-dependent control of melphalan cytotoxicity has been proposed to be responsible for the observed effects [149,150].

The helium-neon laser and other soft lasers have been reported to produce analgesia and wound healing. Studies of laser effects in humans have generally documented decreased pain, inflammation, and oedema in laser-treated tissues. Effects of these lasers on tissues

are biochemical (nonthermal); however, the cellular mechanisms of these effects remains elusive [151,152]. A significant reduction of 5-FU or BMT conditioning regimen-induced oral mucositis has been reported in patients, whose oral mucosa was treated by these lasers [153-156].

4. Systemically applied pharmacotherapeutics

Beta carotene, a vitamin A derivative, is a scavenger of singlet oxygen [157]. It also has significant inhibitory effects on cellular proliferation [158]. In an experimental model, beta carotene supplementation decreased local and systemic toxic effects in irradiated mice [159]. Supplemental dietary beta-carotene led to a mild decrease in the severity of chemotherapy- and radiotherapy-induced oral mucositis in a small study [157].

Azelastine hydrochloride is an antioxidant [160] and a potent histamine H1-receptor antagonist [161]. It suppresses neutrophil respiratory burst both in vivo and in vitro and suppresses cytokine release from lymphocytes [160]. Its prophylactic use reduced the severity of chemoradiotherapy-induced oral mucositis; the result was of little clinical value, however [160].

Pentoxifylline is a xanthine derivative, a haemorrhagic agent, that has been shown to possess profound immunomodulatory properties in vitro, including inhibition of tumor necrosis factor alpha production [162,163]. Elevated levels of tumor necrosis factor alpha have been shown to correlate with both the development and severity of transplantation-related complications [164]. In a phase I-II trial, oral pentoxifylline reduced the frequency and severity of all major complications after BMT, including reduction of oral mucositis severity [165]. However, these results were not reproduced in other studies including the one focused on 5-FU-induced oral mucositis [166-170].

Indomethacin is a nonsteroidal anti-inflammatory drug inhibiting prostaglandin synthesis. It has been reported to delay the onset of mucositis [171].

Treatment with low-dose intramuscular immunoglobulin has been shown to decrease the severity and duration of radiotherapy-induced oral mucositis [172-174]. Prophylactical application of low-dose intramuscular immunoglobulin reduced chemoradiotherapy-induced oral mucositis in patients with head and neck cancer; this reduction lacks clinical relevance, however [175]. Intermediate dose intravenous immunoglobulin G has been observed to prevent high-dose methotrexate-induced oral mucositis [176]. It is probable that anti-inflammatory effects of exogenous immunoglobulin are responsible for the observed effects. Large infusions of immunoglobulin G have been shown to manipulate the immune system; T cell effector and regulatory functions can be manipulated; inflammatory cytokine release

downregulated and complement activation modified [177,178]. In addition, high TGF- β concentrations have been detected in intravenous immunoglobulin preparations [179].

Anticholinergic drugs cause xerostomia by decreasing salivation, which may result in decreased mucosal secretion of certain cytostatic agents and thus reduce their acute toxicity to oral mucosa. Propantheline reduced significantly oral mucositis in patients treated with high-dose etoposide both as single-agent and in multidrug regimen [180,181]. Atropine has not been shown to be beneficial in the prevention of high-dose methotrexate-induced oral mucositis [132].

The mucosal protection effects of granulocyte colony-stimulating factor G-CSF or GM-CSF were observed in patients treated with various chemotherapy regimens [23,182–185], although controversies exist in other clinical trials [186–188]. These studies focused primarily on the myelocytic recovery effect of CSFs on intensive chemotherapy-induced neutropenia. Results of studies evaluating the effect of these cytokines on oral mucositis have confirmed that they are beneficial in reduction of this complication [16,25,189–192]. There are two theories on the mechanism of oral mucositis reduction by G-CSF and GM-CSF. The first one supposes that neutropenia may predispose the patient to oral infections, which may aggravate oral mucositis. G-CSF or GM-CSF may reduce oral mucositis by accelerating neutrophil recovery. The second, more likely mechanism may be a direct stimulative effect of G-CSF or GM-CSF on the growth or regeneration of oral mucosa cells [16,23,25].

Recombinant human interleukin-11 is a pleiotropic cytokine that stimulates bone marrow stem cells to proliferate and exerts effects on the gastrointestinal mucosa which ameliorate responses to injurious stimuli [193]. It has been reported to favorably modify the course of oral mucositis following 5-FU in hamsters [193–195]. This seems to be mediated at the epithelial or connective tissue level rather than through the bone marrow [194].

Reactivation of oral herpes simplex virus is very common in patients receiving cytotoxic chemotherapy or BMT; its incidence rate ranges between 50 and 90% [15,196–199]. However, herpes simplex virus is probably not the major etiologic agent in chemotherapy- and radiotherapy-induced oral mucositis. The incidence of ulcerative mucositis in BMT patients was observed to be high despite the absence of herpes simplex virus and patients who were seronegative were just as likely to develop mucositis as patients who were seropositive, with comparable healing time of ulcers [15,200–202]. Although *acyclovir* prophylaxis is effective in preventing oropharyngeal shedding of the virus in herpes simplex virus seropositive patients receiving intensive chemotherapy or BMT [197,199,201,202], it did not influence

chemotherapy-, radiotherapy-, and BMT-related oral toxicity [201,203–205].

Glutamine is an amino acid synthesized by virtually all tissues and it is an important metabolic substrate for rapidly replicating cells, particularly gastrointestinal tract mucosa and immune cells [206]. During episodes of catabolic stress there is a marked intracellular depletion of glutamine [207]. Parenteral glutamine supplementation appeared to maintain gut integrity in catabolic states in both animal and human studies [208–210] and benefits have also been observed with its enteral administration in methotrexate-related intestinal mucosa toxicity [211]. Oral glutamine supplementation decreased chemotherapy-induced oral mucositis [212,213]; however, no significant positive clinical effect of parenteral or oral glutamine has been observed in two other studies [214,215].

The pineal hormone melatonin inhibits the production of free radicals that mediate the toxicity of chemotherapy. Experimental data have suggested that it may counteract chemotherapy-induced toxicity. Chemotherapy-induced stomatitis was not reduced in a study, although other toxic effects were decreased [216].

5. Other methods

Modification of the chemotherapy regimen may lower its oral toxicity. In the relapsed childhood acute lymphocytic leukemia study of ALL-REZ BFM (Berlin–Frankfurt–Münster)-85, exposition time to methotrexate was observed to be an important factor of mucosa toxicity. Mucosa toxicity was higher in patients treated by intermediate-dose methotrexate in a 36-h infusion than in patients treated by high-dose methotrexate in a 4-h infusion. The end therapy result was similar in both groups [217]. Similar observation of the influence of exposition time on oral mucosa toxicity was made in patients with nasopharyngeal carcinoma treated by cisplatin, 5-FU, and leucovorin (PFL) combination chemotherapy. Using a weekly 24-h infusion schedule of PFL chemotherapy eliminated severe grades of oral mucositis compared to the 5- or 4-day continuous infusion forth- or third-weekly. The significant anticancer activity was retained in the new weekly regimen [26].

The toxic effects of cancer chemotherapy vary according to dosing time because of the effects of circadian rhythms on cellular or proliferative activity [218]. In human oral mucosa, as in other tissues, DNA synthesis, a stage of cell division cycle associated with increased susceptibility to S-phase-specific agents, decreases by 50% or more between 00:00 and 04:00 compared with daytime [218–220]. Severe mucositis occurred in about five times more patients and about 10 times more

courses in patients with metastatic colorectal cancer treated by constant rate infusion of oxaliplatin, FU, and folinic acid than in patients treated by a *chronomodulated infusion* (i.e. oxaliplatin administered during the day hours and FU and folinic acid administered during the night and morning hours). The antitumor effect was greater in the chronomodulated group [221].

6. Conclusion

The long list of solutions, drugs and methods used and studied in the prophylaxis and therapy of chemotherapy- or radiotherapy-induced oral mucositis reflects the need of new, more efficient tools in the management of this complication. Many studies involve only small numbers of patients, which militates against the statistical validity of the reported results that must, therefore, be interpreted with caution. Large studies designed to detect substantial clinical differences are often absent. At the present time, no agent has been shown to be uniformly efficacious and can be accepted as standard therapy. Results of national surveys showed great diversity of mucositis management practices, many of which lack proven clinical efficacy [222-224].

Dental restoration and oral hygiene are basic measures. Oral cooling is a cheap and available method to lower the severity of bolus 5-FU- and melphalan-induced oral mucositis; however, more effective methods are needed. The results of the studies with GM-CSF or G-CSF are promising. Lasers have been shown to be partly beneficial; however, a disadvantage of this method is being equipment-demanding. The modification of the chemotherapy regimen resulting in shortening of the exposition time to chemotherapy agents lowers mucosal toxicity of cisplatin, 5-FU, and leucovorin combination chemotherapy, of high-dose methotrexate, and maybe other regimens. Chronomodulation of chemotherapy might also be helpful. The results of animal studies with locally applied TGF- β 3 and interleukin-11 are also promising. Based on the findings of the role of the inflammatory cascade involved in normal tissues response to chemotherapy and irradiation, further studies should focus on the use of anti-inflammatory drugs. Thalidomide, a drug with immune-modulating activities, which has been reported to be effective in the treatment of aphthous ulcerations of the mouth and oropharynx in patients with human immunodeficiency virus infection, may also deserve study in the management of anticancer therapy-induced oral mucositis [225].

Systematic prospectively designed investigations are necessary in order to achieve a further reduction in the radiotherapy- and chemotherapy-related acute morbidity [224].

References

- [1] National Cancer Institute US Monographs. Consensus Development Conference on Oral Complications of Cancer Therapy: Diagnosis, Prevention, and Treatment. Bethesda: National Institutes of Health, 1990.
- [2] Gallagher JG. Mucositis. In: Klastersky J, Schimpff SC, Lenn HJ, editors. Handbook of Supportive Care in Cancer. New York: Marcel Dekker, 1995. p. 147-56.
- [3] Overholser CD, Jr. Oral care for the cancer patient. In: Klastersky J, Schimpff SC, Lenn HJ, editors. Handbook of Supportive Care in Cancer. New York: Marcel Dekker, 1993. p. 125-45.
- [4] Sonis ST. Oral complications of cancer therapy. In: DeVita VT, Hellma S, Rosenberg SA, editors. Cancer: Principles and Practice of Oncology, 5th Edition. Philadelphia: JB Lippincott, 1993. p. 2385-93.
- [5] Sonis ST. Mucositis as a biological process: a new hypothesis for the development of chemotherapy-induced stomatotoxicity. Oral Oncol 1998;34:39-43.
- [6] Hallahan DE, Haimovitz-Friedman A, Kufe DW, Fuks Z, Weichselbaum RR. The role of cytokines in radiation oncology. Important Adv Oncol 1993;71-80.
- [7] Hallahan D, Kuchibhotla J, Wyble C. Cell adhesion molecules mediate radiation-induced leukocyte adhesion to the vascular endothelium. Cancer Res 1996;56:5150-5.
- [8] Eissner G, Lindner H, Behrends U, Kolch W, Hieke A, Klauke I, Bornkamm GW, Holler E. Influence of bacterial endotoxin on radiation-induced activation of human endothelial cells in vitro and in vivo: protective role of IL-10. Transplantation 1996;62:819-27.
- [9] Koj A. Initiation of acute phase response and synthesis of cytokines. Biochim Biophys Acta 1996;1317:84-94.
- [10] Fisher DE. Apoptosis in cancer therapy: crossing the threshold. Cell 1994;78:539-42.
- [11] Meyn RE, Stephens LC, Milas L. Programmed cell death and radioresistance. Cancer Metastasis Rev 1996;15:119-31.
- [12] Debatin KM. Cytotoxic drugs, programmed cell death, and the immune system: defining new roles in an old play. J Natl Cancer Inst 1997;89:750-1.
- [13] Berkowitz RJ, Crock J, Strickland R, Gordon EM, Strandjord S, Coccia PF. Oral complications associated with bone marrow transplantation in a pediatric population. Am J Pediatr Hematol Oncol 1983;5:53-7.
- [14] Woo SB, Sonis T, Monopoli MM, Sonis AL. A longitudinal study of oral ulcerative mucositis in bone marrow transplant recipients. Cancer 1993;72:1612-7.
- [15] Carrega G, Castagnola E, Canessa A, Argenta P, Haupt R, Dini G, Garaventa A. Herpes simplex virus and oral mucositis in children with cancer. Support Care Cancer 1994;2:266-9.
- [16] Chi KH, Chen CH, Chan WK, Chow KC, Chen SY, Yen SH, Chao JY, Chang CY, Chen KY. Effect of granulocyte-macrophage colony-stimulating factor on oral mucositis in head and neck cancer patients after cisplatin, fluorouracil, and leucovorin chemotherapy. J Clin Oncol 1995;13:2620-8.
- [17] Rosenberg S. Oral complications of cancer chemotherapy—a review of 398 patients. J Oral Med 1986;41:93-7.
- [18] Hasenau C, Clasen BPE, Roettger D. Anwendung einer standardisierten Mundpflege zur Prophylaxe und Therapie einer Mukositis bei Patienten während der Radiochemotherapie von Kopf-Hals-Malignomen. Laryng Rhinol Otol 1988; 67:576-9.
- [19] Mailath G, Rasse M, Rotter M, Hollmann K. Zur Ursache der Stomatitis bei der Chemotherapie von Kopf-Hals-Tumoren. Z Stomatol 1989;86:353-9.
- [20] Spijkervet FKL, van Saene HKF, van Saene JJM, Panders AK, Vermey A, Mehta DM, Fidler V. Effect of selective

- elimination of the oral flora on mucositis in irradiated head and neck cancer patients. *J Surg Oncol* 1991; 46:167-73.
- [1] Martin MV. Irradiation mucositis: a reappraisal. *Oral Oncol Eur J Cancer* 1993;29B:1-2.
 - [2] Symonds RP, McIlroy P, Khorrami J, Paul J, Pyper E, Alcock SR, McCallum I, Spekenbrink AB, McMurray A, Lindemann E, Thomas M. The reduction of radiation mucositis by selective decontamination antibiotic pastilles: a placebo-controlled double-blind trial. *Br J Cancer* 1996;74:312-7.
 - [3] Gabrilove JL, Jakubowski A, Scher H, Sternberg C, Wong G, Grous J, Yagoda A, Fain K, Moore MAS, Clarkson B, Oettingen HF, Alton K, Welte K, Souza L. Effect of granulocyte colony-stimulating factor on neutropenia and associated morbidity due to chemotherapy for transitional cell carcinoma of the urothelium. *N Engl J Med* 1988;318:1414-22.
 - [4] LeVeque FG, Parzuchowski JB, Farinacci GC, Redding SW, Rodu B, Johnson JT, Ferretti GA, Eisenberg PD, Zimmer MB. Clinical evaluation of MGI 209, an anesthetic, film-forming agent for relief from painful oral ulcers associated with chemotherapy. *J Clin Oncol* 1992;10:1963-8.
 - [5] Gordon B, Spadinger A, Hodges E, Ruby E, Stanley R, Coccia P. Effect of granulocyte-macrophage colony-stimulating factor on oral mucositis after hematopoietic stem-cell transplantation. *J Clin Oncol* 1994;12:1917-22.
 - [6] Chi KH, Chan WK, Shu CH, Law CK, Chen SY, Yen SH, Chen KY. Elimination of dose limiting toxicities of cisplatin, 5-fluorouracil, and leucovorin using a weekly 24-hour infusion schedule for the treatment of patients with nasopharyngeal carcinoma. *Cancer* 1995;76:2186-92.
 - [7] Ibrahim EM, al-Mulhim FA. Effect of granulocyte-macrophage colony-stimulating factor on chemotherapy-induced oral mucositis in non-neutropenic cancer patients. *Med Oncol* 1997;14:47-51.
 - [8] Weisdorf DJ, Bostrom B, Raether D, Mattingly M, Walker P, Pihlstrom B, Ferrieri P, Haake R, Goldman A, Woods W, Ramsay NKC, Kersey JH. Oropharyngeal mucositis complicating bone marrow transplantation: prognostic factors and the effect of chlorhexidine mouth rinse. *Bone Marrow Transplant* 1989;4:89-95.
 - [9] Spijkervet FKL, van Saene HKF, Panders AK, Vermey A, van Saene JJM, Mehta DM, Fidler V. Effect of chlorhexidine rinsing on the oropharyngeal ecology in patients with head and neck cancer who have irradiation mucositis. *Oral Surg Oral Med Oral Pathol* 1989;67:154-61.
 - [10] Epstein JB, Vickars L, Spinelli J, Reece D. Efficacy of chlorhexidine and nystatin rinses in prevention of oral complications in leukemia and bone marrow transplantation. *Oral Surg Oral Med Oral Pathol* 1992;73:682-9.
 - [11] Foote RL, Loprinzi CL, Frank AR, O'Fallon JR, Gulavita S, Tewfik HH, Ryan MA, Earle JM, Novotny P. Randomized trial of a chlorhexidine mouthwash for alleviation of radiation-induced mucositis. *J Clin Oncol* 1994;12:2630-3.
 - [12] Bondi E, Baroni C, Prete A, Gatti M, Carrassi A, Lodi G, Porter SR. Local antimicrobial therapy of oral mucositis in paediatric patients undergoing bone marrow transplantation. *Oral Oncol* 1997;33:322-6.
 - [13] Okuno SH, Foote RL, Loprinzi CL, Gulavita S, Sloan JA, Earle J, Novotny PJ, Burk M, Frank AR. A randomized trial of a nonabsorbable antibiotic lozenge given to alleviate radiation-induced mucositis. *Cancer* 1997;79:2193-9.
 - [14] Wilkes JD. Prevention and treatment of oral mucositis following cancer chemotherapy. *Semin Oncol* 1998;25:538-51.
 - [15] Wright WE, Haller JM, Harlow SA, Pizzo PA. An oral disease prevention program for patients receiving radiation and chemotherapy. *J Am Dent Assoc* 1985;110:43-7.
 - [16] Sonis S, Kunz A. Impact of improved dental services on the frequency of oral complications of cancer therapy for patients with non-head-and-neck malignancies. *Oral Surg Oral Med Oral Pathol* 1988;65:19-22.
 - [17] National Institutes of Health. National Institutes of Health consensus development conference statement: oral complications of cancer therapies: diagnosis, prevention, and treatment. *J Am Dent Assoc* 1989;119:179-83.
 - [18] Jansma J, Vissink A, Spijkervet FKL, Roodenburg JLN, Panders AK, Vermey A, Szabó BG, Gravenmade EJ. Protocol for the prevention and treatment of oral sequelae resulting from head and neck radiation therapy. *Cancer* 1992;70:2171-80.
 - [19] Whitmyer CC, Esposito SJ, Terezhalmy GT. Radiotherapy for head and neck neoplasms. *Gen Dent* 1997;45:363-70.
 - [20] Borowski B, Benhamou E, Pico JL, Laplanche A, Marginaud JP, Hayat M. Prevention of oral mucositis in patients treated with high-dose chemotherapy and bone marrow transplantation: a randomised controlled trial comparing two protocols of dental care. *Eur J Cancer B Oral Oncol* 1994;30B:93-7.
 - [21] Carl W, Emrich LS. Management of oral mucositis during local radiation and systemic chemotherapy: a study of 98 patients. *J Prosthet Dent* 1991;66:361-9.
 - [22] Bernhof CH, Skaug N. Oral findings in irradiated edentulous patients. *Int J Oral Surg* 1985;14:416-27.
 - [23] Carl W. Oral complications of local and systemic cancer treatment. *Curr Opin Oncol* 1995;7:320-4.
 - [24] Feber T. Management of mucositis in oral irradiation. *Clin Oncol R Coll Radiol* 1996;8:106-11.
 - [25] Shieh SH, Wang ST, Tsai ST, Tseng CC. Mouth care for nasopharyngeal cancer patients undergoing radiotherapy. *Oral Oncol* 1997;33:36-41.
 - [26] Richardson CT. Sucralfate. *Ann Intern Med* 1982;97:269-72.
 - [27] Nagashima R, Yoshida N, Terao N. Sucralfate, a basic aluminum salt of sucrose sulfate. II. Inhibition of peptic hydrolysis as it results from sucrose sulfate interaction with protein substrate, serum albumins. *Arzneimittelforschung* 1979;29:73-6.
 - [28] Nagashima R, Hirano T. Selective binding of sucralfate to ulcer lesion. I. Experiments in rats with acetic acid-induced gastric ulcer receiving unlabelled sucralfate. *Arzneimittelforschung* 1980;30:80-3.
 - [29] Szabo S, Hollander D. Pathways of gastrointestinal protection and repair: mechanisms of action of sucralfate. *Am J Med* 1989;86:23-31.
 - [30] Szabo S, Vattay P, Scarbrough E, Folkman J. Role of vascular factors including angiogenesis in the mechanisms of action of sucralfate. *Am J Med* 1991;91:158-60.
 - [31] Ferraro JM, Mattern JQA, II. Sucralfate suspension for stomatitis. *Drug Intell Clin Pharm* 1984;18:153 (letter).
 - [32] Ferraro JM. Sucralfate suspension for mouth ulcers. *Drug Intell Clin Pharm* 1985;19:480 (letter).
 - [33] Solomon MA. Oral sucralfate suspension for mucositis. *N Engl J Med* 1986;315:459-60 (letter).
 - [34] Pfeiffer P, Madsen EL, Hansen O, May O. Effect of prophylactic sucralfate suspension on stomatitis induced by cancer chemotherapy. A randomized, double-blind cross-over study. *Acta Oncol* 1992;31:171-3.
 - [35] Scherlacher A, Beaufort-Spontan F. Strahlentherapie von Kopf-Hals-Malignomen: Entzündungsprophylaxe der Schleimhaut durch Sucralfatbehandlung. *HNO* 1990;38:24-8.
 - [36] Allison RR, Vongtama V, Vaughan J, Shin KH. Symptomatic acute mucositis can be minimized or prophylaxed by the combination of sucralfate and fluconazole. *Cancer Invest* 1995;13:16-22.
 - [37] Franzén L, Henriksson R, Littbrand B, Zackrisson B. Effects of sucralfate on mucositis during and following radiotherapy of malignancies in the head and neck region. *Acta Oncologica* 1995;34:219-23.
 - [38] Giorgi F, Bascioni R, De Signoribus G, Di-Saverio F. Sucralfate prophylaxis of fluorouracil-induced stomatitis. *Tumori* 1996;82:585-7.

- [59] Shenep JL, Kalwinsky DK, Hutson PR, George SL, Dodge RK, Blankenship KR, Thornton D. Efficacy of oral sucralfate suspension in prevention and treatment of chemotherapy-induced mucositis. *J Pediatr* 1988;113:758-63.
- [60] Epstein JB, Wong FL. The efficacy of sucralfate suspension in the prevention of oral mucositis due to radiation therapy. *Int J Radiat Oncol Biol Phys* 1994;28:693-8.
- [61] Makkonen TA, Bostrom P, Vilja P, Joensuu H. Sucralfate mouth washing in the prevention of radiation-induced mucositis: a placebo-controlled double-blind randomized study. *Int J Radiat Oncol Biol Phys* 1994;30:177-82.
- [62] Loprinzi CL, Ghosh C, Camoriano J, Sloan J, Steen PD, Michalak JC, Schaefer PL, Novotny PJ, Gerstner JB, White DF, Hatfield AK, Quella SK. Phase III controlled evaluation of sucralfate to alleviate stomatitis in patients receiving fluorouracil-based chemotherapy. *J Clin Oncol* 1997; 15:1235-8.
- [63] Meredith R, Salter M, Kim R, Spencer S, Weppelmann B, Rodu B, Smith J, Lee J. Sucralfate for radiation mucositis: results of a double-blind randomized trial. *Int J Radiat Oncol Biol Phys* 1997;37:275-9.
- [64] Barker G, Loftus L, Cuddy P, Barker B. The effects of sucralfate suspension and dihenhydramine syrup plus kaolin-pectin on radiotherapy-induced mucositis. *Oral Surg Oral Med Oral Pathol* 1991;71:288-93.
- [65] Tampo Y, Yonaha M. Vitamin E and glutathione are required for preservation of microsomal glutathione s-transferase from oxidative stress in microsomes. *Pharmacol Toxicol* 1990; 66:259-65.
- [66] Kagan V, Serbinova E, Packer L. Antioxidant effects of ubiquinones in microsomes and mitochondria are mediated by tocopherol recycling. *Biochem Biophys Res Commun* 1990; 169:851-7.
- [67] Wilson JD. Vitamin deficiency and excess. In: Wilson JD, Braunwald E, Isselbacher KJ, Petersdorf RG, Martin JB, Fauci AS, Root RK, editors. *Harrison's Principles of Internal Medicine*, 12th Edition. New York: McGraw-Hill, 1991. p. 434-45.
- [68] Schreck R, Rieber P, Baeuerle PA. Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF- κ B transcription factor and HIV-1. *EMBO J* 1991;10:2247-58.
- [69] Wadleigh RG, Redman RS, Graham ML, Krasnow SH, Anderson A, Cohen MH. Vitamin E in the treatment of chemotherapy-induced mucositis. *Am J Med* 1992;92:481-4.
- [70] Lopez I, Goudou C, Ribrag V, Sauvage C, Hazebrucq G, Dreyfus F. Traitement des mucites par la vitamine E lors de l'administration d'anti-neoplastiques neutropénisants. *Ann Med Interne Paris* 1994;145:405-8.
- [71] Prutkin L. Wound healing and vitamin A acid. *Acta Dermatol Venereol* 1972;52:489-92.
- [72] Orfanos CE, Bauer R. Evidence for anti-inflammatory activities of oral synthetic retinoids: experimental findings and clinical experience. *Br J Dermatol* 1983;109:55-60.
- [73] Sporn MB, Roberts AB. Regulation of cell differentiation and proliferation by retinoids and transforming growth factor β . In: Burger MM, Sordat B, Zinkenagel RM, editors. *Cell to Cell Interaction*. Basel: Karger, 1990. p. 2-15.
- [74] Cohen G, Elad S, Or R, Galili D, Garfunkel AA. The use of tretinoin as oral mucositis prophylaxis in bone marrow transplantation patients: a preliminary study. *Oral Dis* 1997;3:243-6.
- [75] Daeflfer R. Oral hygiene measures for patients with cancer. *Cancer Nurs* 1980;3:347-56.
- [76] Dudjak LA. Mouth care for mucositis due to radiation therapy. *Cancer Nurs* 1987;10:131-40.
- [77] McGaw WT, Belch A. Oral complications of acute leukemia: prophylactic impact of chlorhexidine mouth rinse regimen. *Oral Surg Oral Med Oral Pathol* 1985;60:275-80.
- [78] Ferretti GA, Ash RC, Brown AT, Parr MD, Romond EH, Lillich TT. Control of oral mucositis and candidiasis in marrow transplantation: a prospective, double-blind trial of chlorhexidine digluconate oral rinse. *Bone Marrow Transplant* 1988; 3:483-93.
- [79] Ferretti GA, Raybould TP, Brown AT, MacDonald JS, Greenwood M, Maruyama Y, Geil J, Lillich TT, Ash RC. Chlorhexidine prophylaxis for chemotherapy- and radiotherapy-induced stomatitis: a randomized double-blind trial. *Oral Surg Oral Med Oral Pathol* 1990;69:331-8.
- [80] Rutkauskas JS, Davis JW. Effects of chlorhexidine during immunosuppressive chemotherapy: a preliminary report. *Oral Surg Oral Med Oral Pathol* 1993;76:441-8.
- [81] Wahlin BY. Effects of chlorhexidine mouthrinse on oral health in patients with acute leukemia. *Oral Surg Oral Med Oral Pathol* 1989;68:279-87.
- [82] Dodd MJ, Larson PJ, Dibble SL, Miaskowski C, Greenspan D, MacPhail L, Hauck WW, Paul SM, Ignoffo R, Shiba G. Randomized clinical trial of chlorhexidine versus placebo for prevention of oral mucositis in patients receiving chemotherapy. *Oncol Nurs Forum* 1996;23:921-7.
- [83] Rahn R, Adamietz IA, Böttcher HD, Schäfer V, Reimer K, Fleischer W. Povidone-iodine to prevent mucositis in patients during antineoplastic radiochemotherapy. *Dermatology* 1997; 195 (Suppl. 2):57-61.
- [84] Martin MV, Al-Tikriti U, Bramley PA. Yeast flora of the mouth and skin during and after irradiation for oral and laryngeal cancer. *J Med Microbiol* 1981;14:457-67.
- [85] Pau HW, Strähler-Pohl HJ, Exner M. Hefepilzflora bei der Radiotherapie von Tumoren den oberen Aerodigestiven Trakt. *HNO* 1985;33:485-8.
- [86] Wade JC, Schimpff SC. Epidemiology and prevention of Candida infection. In: Bodey GP, Famstein V, editors. *Candidosis*. New York: Raven Press, 1985. p. 111-13.
- [87] Bernhoft CH, Skaug N. Oral findings in irradiated edentulous patients. *Int J Oral Surg* 1985;14:416-27.
- [88] Ferretti GA, Brown AT, Lillich TT, Ash RC, Largent BM. The effect of chlorhexidine on the oral microflora of bone marrow transplant patients. *J Dent Res* 1985;64 (Suppl. 6):235.
- [89] Matthews RH, Ercal N. Prevention of mucositis in irradiated head and neck cancer patients. *J Exp Ther Oncol* 1996;1:135-8.
- [90] Thieme K, Stadler R, Isaac O. Biochemical investigations of the camomile. *Arzneimittelforschung* 1973;23:756-9.
- [91] Fidler P, Loprinzi CL, O'Fallon JR, Leitch JM, Lee JK, Hayes DL, Novotny P, Clemens-Schutjer D, Bartel J, Michalak JC. Prospective evaluation of a chamomile mouthwash for prevention of 5-FU-induced oral mucositis. *Cancer* 1996;77:522-5.
- [92] Scheinman RI, Gualberto A, Jewell CM, Cidlowski JA, Baldwin AS, Jr. Characterization of mechanisms involved in transrepression of NF- κ B by activated glucocorticoid receptors. *Mol Cell Biol* 1995;15:943-53.
- [93] Barnes PJ, Karin M. Nuclear factor- κ B—a pivotal transcription factor in chronic inflammatory diseases. *N Engl J Med* 1997;336:1066-71.
- [94] Abdelaal AS, Barker DS, Fergusson MM. Treatment for irradiation-induced mucositis. *Lancet* 1989;1:97 (letter).
- [95] Hunter KM. A clinical evaluation of benzydamine hydrochloride. *Aust Dent J* 1978;23:164-6.
- [96] Kataoka S, Nishimura K, Naito T. In vivo metabolism and anti-inflammatory activity of benzydamine hydrochloride in rats treated with carrageenin. *Chem Pharm Bull (Tokyo)* 1979;27:2890-903.
- [97] Whiteside MW. A controlled study of benzydamine oral rinse ("Difflam") in general practice. *Curr Med Res Opin* 1982; 8:188-90.
- [98] Kim JH, Chu FC, Lakshmi V, Houde R. Benzydamine HCl, a new agent for the treatment of radiation mucositis of the oropharynx. *Am J Clin Oncol* 1986;9:132-4.

- [99] Prada A, Chiesa F. Effects of benzydamine on the oral mucositis during antineoplastic radiotherapy and/or intra-arterial chemotherapy. *Int J Tiss Reac* 1987;9:115-9.
- [100] Epstein JB, Stevenson-Moore P, Jackson S, Mohamed JH, Spinelli JJ. Prevention of oral mucositis in radiation therapy: a controlled study with benzydamine hydrochloride rinse. *Int J Radiat Oncol Biol Phys* 1989;16:1571-5.
- [101] Samaranyake LP, Robertson AG, MacFarlane TW, Hunter IP, MacFarlane G, Soutar DS, Ferguson MM. The effect of chlorhexidine and benzydamine mouthwashes on mucositis induced by therapeutic irradiation. *Clin Radiol* 1988;39:291-4.
- [102] Schubert MM, Newton RE. The use of benzydamine HCl for the management of cancer therapy-induced mucositis: preliminary report of a multicentre study. *Int J Tissue Reac* 1987;9:99-103.
- [103] Epstein JB, Stevenson-Moore P. Benzydamine hydrochloride in prevention and management of pain in oral mucositis associated with radiation therapy. *Oral Surg Oral Med Oral Pathol* 1986;62:145-8.
- [104] Lever SA, Dupuis LL, Chan SL. Comparative evaluation of benzydamine oral rinse in children with antineoplastic-induced stomatitis. *Drug Intell Clin Pharm* 1987;21:359-61.
- [105] Glickman RM. Inflammatory bowel disease. Ulcerative colitis and Crohn's disease. In: Wilson JD, Braunwald E, Isselbacher KJ, Petersdorf RG, Martin JB, Fauci AS, Root RK, editors. *Harrison's Principles of Internal Medicine*, II, 12th Edition. New York: McGraw-Hill, 1991. p. 1268-81.
- [106] Kopp E, Ghosh S. Inhibition of NF- κ B by sodium salicylate and aspirin. *Science* 1994;265:956-9.
- [107] Collier PM, Neill SM, Copeman PW. Topical 5-aminosalicylic acid: a treatment for aphthous ulcers. *Br J Dermatol* 1992;126:185-8.
- [108] Ranzi T, Campanini M, Bianchi P. Successful treatment of genital and oral ulceration in Behcet's disease with topical 5-aminosalicylic acid (5-ASA). *Br J Dermatol* 1989;120:471 (letter).
- [109] Rymes N, Glick L, Holmes JA. Topical mesalazine in the treatment of chemotherapy and radiotherapy-induced oral mucositis. *Bone Marrow Transplant* 1996;18:484 (letter).
- [110] Cartee L, Petros WP, Rosner GL, Gilbert C, Moore S, Affronti ML, Hoke JA, Hussein AM, Ross M, Rubin P, Vredenburg JJ, Peters WP. Evaluation of GM-CSF for prevention of chemotherapy-induced mucositis: a randomized, double-blind, dose-ranging study. *Cytokine* 1995; 7:471-7.
- [111] Cox DA, Maurer T. Transforming growth factor- β . *Clin Immunol Immunopathol* 1997;83:25-30.
- [112] Sonis ST, Lindquist L, van Vugt V, Stewart AA, Stam K, Qu GY, Iwata KK, Haley JD. Prevention of chemotherapy-induced ulcerative mucositis by transforming growth factor β 3. *Cancer Res* 1994;54:1135-8.
- [113] Sonis ST, van Vugt AG, Brien JP, Muska AD, Bruskin AM, Rose A, Haley JD. Transforming growth factor- β 3 mediated modulation of cell cycling and attenuation of 5-fluorouracil induced oral mucositis. *Oral Oncol* 1997; 33:47-54.
- [114] Cohen S. The epidermal growth factor (EGF). *Cancer* 1983; 51:1787-91.
- [115] Epstein JB, Emerton S, Guglietta A, Le N. Assessment of epidermal growth factor in oral secretions of patients receiving radiation therapy for cancer. *Oral Oncol* 1997;33:359-63.
- [116] Girdler NM, McGurk M, Aqual S, Prince M. The effect of epidermal growth factor mouthwash on cytotoxic-induced oral ulceration. A phase I clinical trial. *Am J Clin Oncol* 1995; 18:403-6.
- [117] Pastuszak AL, Schöler L, Speck-Martins CE, Coelho KEFA, Cordello SM, Vargas F, Brunoni D, Schwarz IVD, Larranda-baru M, Safatle H, Meloni VFA, Koren G. Use of misoprostol during pregnancy and Möbius' syndrome in infants. *N Engl J Med* 1998;338:1881-5.
- [118] Duenas-Gonzalez A, Sobrevilla-Calvo P, Frias-Medivil M, Gallardo-Rincon D, Lara-Medina F, Aguilar-Ponce L, Miranda-Lopez E, Zinser-Sierra J, Reynoso-Gomez E. Misoprostol prophylaxis for high-dose chemotherapy-induced mucositis: a randomized double-blind study. *Bone Marrow Transplant* 1996;17:809-12.
- [119] Johansson C, Bergstrom S. Prostaglandins and protection of gastroduodenal mucosa. *Scand J Gastroenterol* 1982;77:21-46.
- [120] Kühner I, Kuzmits R, Linkesch W, Ludwig H. Topical PGE₂ enhances healing of chemotherapy associated mucosal lesions. *Lancet* 1986;1:622 (letter).
- [121] Porteder H, Rausch E, Kment G, Watzek G, Matejka M, Sinzinger H. Local prostaglandin E₂ in patients with oral malignancies undergoing chemo- and radiotherapy. *J Cranio-maxillofac Surg* 1988;16:371-4.
- [122] Matejka M, Nell A, Kment G, Schein A, Leukauf M, Porteder H, Mailath G, Sinzinger H. Local benefit of prostaglandin E₂ in radiochemotherapy-induced oral mucositis. *Br J Oral Maxillofac Surg* 1990;28:89-91.
- [123] Labar B, Mršić M, Pavletić Ž, Bogdanić V, Nemet D, Aurer I, Radman I, Filipovic-Grcić N, Sertić D, Kalenić S, Presečki V. Prostaglandin E₂ for prophylaxis of oral mucositis following BMT. *Bone Marrow Transplant* 1993;11:379-82.
- [124] Fiocchi C. Cytokines and intestinal inflammation. *Transplant Proc* 1996;28:2442-3.
- [125] Czusak CA, Sutherland DE, Billman MA, Stein SH. Prostaglandin E₂ potentiates interleukin-1 beta induced interleukin-6 production by human gingival fibroblasts. *J Clin Periodontol* 1996;23:635-40.
- [126] Rothwell BR, Spektor WS. Palliation of radiation-related mucositis. *Spec Care Dentist* 1990;10:21-5.
- [127] Simon AR, Roberts MW. Management of oral complications associated with cancer therapy in pediatric patients. *ASDC J Dent Child* 1991;58:384-9.
- [128] Maciejewski B, Zajusz A, Pilecki B, Swiatnicka J, Składowski K, Dorr W, Kummermehr J, Trott KF. Acute mucositis in the stimulated oral mucosa of patients during radiotherapy for head and neck cancer. *Radiother Oncol* 1991;22:7-11.
- [129] Maciejewski B, Składowski K, Zajusz A. Radiobiological predictors of tumor and acute normal tissue response in radiotherapy for head and neck cancers. *Neoplasma* 1991; 38:513-22.
- [130] Dorr W, Jacubek A, Kummermehr J, Herrmann T, Dolling-Jochem I, Eckelt U. Effects of stimulated repopulation on oral mucositis during conventional radiotherapy. *Radiother Oncol* 1995;37:100-7.
- [131] Ackland SP, Schilsky RL. High-dose methotrexate: a critical reappraisal. *J Clin Oncol* 1987;5:2017-31.
- [132] Oliff A, Bleyer WA, Poplack DG. Methotrexate-induced oral mucositis and salivary methotrexate concentrations. *Cancer Chemother Pharmacol* 1979;2:225-6.
- [133] Rask C, Albertioni F, Schröder H, Peterson C. Oral mucositis in children with acute lymphoblastic leukemia after high-dose methotrexate treatment without delayed elimination of methotrexate: relation to pharmacokinetic parameters of methotrexate. *Pediatr Hematol Oncol* 1996;13:359-67.
- [134] Albertioni F, Rask C, Schroeder H, Peterson C. Monitoring of methotrexate 7-hydroxymethotrexate in saliva from children with acute lymphoblastic leukemia receiving high-dose consolidation treatment: relation to oral mucositis. *Anticancer Drugs* 1997;8:119-24.
- [135] Fox RM, Woods RL, Tattersall MHN, Brodie GM. Allopurinol modulation of high-dose fluorouracil toxicity. *Lancet* 1979;1:677 (letter).
- [136] Schwartz PM, Dunigan JM, Marsh JC, Handschumacher RE. Allopurinol modification of the toxicity and antitumor activity of 5-fluorouracil. *Cancer Res* 1980;40:1885-9.

- [137] Howell SB, Wung WE, Taetle R, Hussain F, Romine JS. Modulation of 5-fluorouracil toxicity by allopurinol in man. *Cancer* 1981;48:1281-9.
- [138] Nakamura K, Natsugoe S, Kumano T, Shinkawa T, Kariyazono H, Yamada K, Baba M, Yoshinaka H, Fukumoto T, Aikou T. Prophylactic action of allopurinol against chemotherapy-induced stomatitis—inhibition of superoxide dismutase and proteases. *Anticancer Drugs* 1996;7:235-9.
- [139] Clark PI, Slevin ML. Allopurinol mouthwashes and 5-fluorouracil induced oral toxicity. *Eur J Surg Oncol* 1985;11:267-8.
- [140] Tsavaris N, Caragiannis P, Kosmidis P. Reduction of oral toxicity of 5-fluorouracil by allopurinol mouthwashes. *Eur J Surg Oncol* 1988;14:405-6.
- [141] Porta C, Moroni M, Nastasi G. Allopurinol mouthwashes in the treatment of 5-fluorouracil-induced stomatitis. *Am J Clin Oncol* 1994;17:246-7.
- [142] Loprinzi CL, Cianflone SG, Dose AM, Etzell PS, Burnham NL, Therneau TM, Hagen L, Gainey DK, Cross M, Athmann LM, Fischer T, O'Connell MJ. A controlled evaluation of an allopurinol mouthwash as prophylaxis against 5-fluorouracil-induced stomatitis. *Cancer* 1990;65:1879-82.
- [143] New shield protects oral tissues during radiation. *J Am Dent Assoc* 1994;125:508 (news).
- [144] Perch SJ, Machtay M, Markiewicz DA, Klingerman MM. Decreased acute toxicity by using midline mucosa-sparing blocks during radiation therapy for carcinoma of the oral cavity, oropharynx, and nasopharynx. *Radiology* 1995;197:863-6.
- [145] Mahood DJ, Dose AM, Loprinzi CL, Veeder MH, Athmann LM, Therneau TM, Sorenson JM, Gainey DK, Mailliard JA, Gusa NL, Finck GK, Johnson C, Goldberg RM. Inhibition of fluorouracil-induced stomatitis by oral cryotherapy. *J Clin Oncol* 1991;9:449-52.
- [146] Cascinu S, Fedeli A, Fedeli SL, Catalano G. Oral cooling (cryotherapy), an effective treatment for the prevention of 5-fluorouracil-induced stomatitis. *Oral Oncol Eur J Cancer* 1994;30B:234-6.
- [147] Sato A, Kumagai S, Sakaki K, Morikawa H, Song ST, Mori S. Inhibition of 5-fluorouracil-cisplatin-induced stomatitis by oral cryotherapy: use of an ice-bar containing fibrinolysin and deoxyribonuclease combine (Elate). *Gan To Kagaku Ryoho* 1997;24:1135-9.
- [148] Rocke LK, Loprinzi CL, Lee JK, Kunselman SJ, Iverson RK, Finck G, Lifsey D, Glaw KC, Stevens BA, Hatfield AK, Vaught NL, Bartel J, Pierson N. A randomized clinical trial of two different durations of oral cryotherapy for prevention of 5-fluorouracil related stomatitis. *Cancer* 1993;72:2234-8.
- [149] Dumontet C, Sonnet A, Bastion Y, Salles G, Espinouse D, Coiffier B. Prevention of high dose L-PAM-induced mucositis by cryotherapy. *Bone Marrow Transplant* 1994;14:492-4 (letter).
- [150] Meloni G, Capria S, Proia A, Trisolini SM, Mandelli F. Ice pops to prevent melphalan-induced stomatitis. *Lancet* 1996;347:1691-2.
- [151] Basford JR. Low energy laser therapy: controversies and new research findings. *Mayo Clin Proc* 1986;61:671-5.
- [152] King PR. Low level laser therapy: a review. *Laser Med Sci* 1989;4:141-9.
- [153] Ciais G, Namer M, Schneider M, Demard F, Pourreau-Schneider N, Martin PM, Soudry M, Franquin JC, Zattara H. La lasertherapie dans la prevention et le traitement des mucites liees a la chimiotherapie anticanceruse. *Bull Cancer* 1992;79:183-91.
- [154] Pourreau-Schneider N, Soudry M, Franquin JC, Zattara H, Martin PM, Ciais G, Namer M, Schneider M, Chauvel P, Demard F. Soft-laser therapy for iatrogenic mucositis in cancer patients receiving high-dose fluorouracil: a preliminary report. *J Natl Cancer Inst* 1992;84:358-9.
- [155] Barasch A, Peterson DE, Tanzer JM, D'Ambrosio JA, Nuki K, Schubert MM, Franquin JC, Clive J, Tutschka P. Helium-neon laser effects on conditioning-induced oral mucositis in bone marrow transplantation patients. *Cancer* 1995;76:2550-6.
- [156] Cowen D, Tardieu C, Schubert M, Peterson D, Resbeut M, Faucher C, Franquin JC. Low energy Helium-Neon laser in the prevention of oral mucositis in patients undergoing bone marrow transplant: results of a double blind randomized trial. *Int J Radiat Oncol Biol Phys* 1997;38:697-703.
- [157] Mills EED. The modifying effect of beta-carotene on radiation and chemotherapy-induced oral mucositis. *Br J Cancer* 1988;57:416-7.
- [158] Garewal HS, Meyskens FL, Killen D, Reeves D, Kiersch TA, Elletson H, Strosberg A, King D, Steinbronn K. Response of oral leukoplakia to beta-carotene. *J Clin Oncol* 1990;8:1715-20.
- [159] Seifter E, Returra G, Padawer J, Stratford F, Goodwin P, Levenson SM. Regression of C₃HBA mouse tumor due to X-ray therapy combined with supplemental β -carotene or vitamin A. *J Natl Cancer Inst* 1983;71:409-17.
- [160] Osaki T, Ueta E, Yoneda K, Hirota J, Yamamoto T. Prophylaxis of oral mucositis associated with chemoradiotherapy for oral carcinoma by Azelastine hydrochloride (Azelastine) with other antioxidants. *Head Neck* 1994;16:331-9.
- [161] Horak F. Seasonal allergic rhinitis. Newer treatment approaches. *Drugs* 1993;45:518-27.
- [162] Han J, Thompson P, Beutler B. Dexamethasone and pentoxifylline inhibit endotoxin-induced cachectin/tumor necrosis factor synthesis at separate points in the signaling pathway. *J Exp Med* 1990;172:391-4.
- [163] Tilg H, Eibl B, Pichl M, Gachter A, Herold M, Brankova J, Huber C, Niederwieser D. Immune response modulation by pentoxifylline in vitro. *Transplantation* 1993;56:196-201.
- [164] Holler E, Kolb HJ, Möller A, Kempeni J, Liesenfeld S, Pechmer H, Lehmacher W, Ruckdeschel G, Gleixner B, Riedner C. Increased serum levels of tumor necrosis factor alpha precede major complications of bone marrow transplantation. *Blood* 1990;75:1011-16.
- [165] Bianco JA, Appelbaum FR, Nemunaitis J, Almgren J, Andrews F, Kettner P, Shields A, Singer JW. Phase I-II trial of pentoxifylline for the prevention of transplant-related toxicities following bone marrow transplantation. *Blood* 1991;78:1205-11.
- [166] Stockschrader M, Kalhs P, Peters S, Zeller W, Kruger W, Kabisch H, Lechner K, Zander A. Intravenous pentoxifylline failed to prevent transplant-related toxicities in allogeneic bone marrow transplant recipients. *Bone Marrow Transplant* 1993;12:357-62.
- [167] Clift RA, Bianco JA, Appelbaum FR, Buckner CD, Singer JW, Bakke L, Bensinger WI, Bowden RA, McDonald GB, Schubert M, Shields AF, Slattery JT, Storb R, Fisher LD, Mori M, Thomas ED, Hansen JA. A randomized controlled trial of pentoxifylline for the prevention of regimen-related toxicities in patients undergoing allogeneic marrow transplantation. *Blood* 1993;82:2025-30.
- [168] Attal M, Huguet F, Rubie H, Charlet JP, Schlaifer D, Huynh A, Laurent G, Pris J. Prevention of regimen-related toxicities after bone marrow transplantation by pentoxifylline: a prospective, randomized trial. *Blood* 1993;82:732-6.
- [169] van der Jagt RHC, Pari G, McDiarmid SA, Boisvert DM, Huebsch LB. Effect of pentoxifylline on regimen related toxicity in patients undergoing allogeneic or autologous bone marrow transplantation. *Bone Marrow Transplant* 1994;13:203-7.
- [170] Verdi CJ, Garewal HS, Koenig LM, Vaughn B, Burkhead T. A double-blind, randomized, placebo-controlled, crossover trial of pentoxifylline for the prevention of chemotherapy-induced oral mucositis. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1995;80:36-42.

- [171] Pillsbury HC, 3rd, Webster WP, Rosenman JG. Prostaglandin inhibitor and radiotherapy in advanced head and neck cancers. *Arch Otolaryng Head Neck Surg* 1986;112:552-3.
- [172] Schedler M, Feidt H, Niewaldt M. Lässt sich die radiogene Mukositis mit Immunglobulinen behandeln? *Zentralbl Hals Ohren Heilkd Kopf Haschir* 1990;139:3-4.
- [173] Proske H, Pfab R. Immunglobulinpräparate als antiinflammatorische Therapeutika in der Strahlentherapie. *Med Welt* 1992;43:1025-6.
- [174] Schedler MGJ, Bost P, Federspil P, Pautler M, Schätzle W. Die Behandlung der strahleninduzierten Mukositis bei Kopf-/Halstumoren mit intramuskulär verabreichtem polyvalentem Immunglobulin. *Tumor Diagn Ther* 1994;15:184-91.
- [175] Mose S, Adamietz IA, Saran F, Thilmann C, Heyd R, Knecht R, Bottcher HD. Can prophylactic application of immunoglobulin decrease radiotherapy-induced oral mucositis? *Am J Clin Oncol* 1997;20:407-11.
- [176] Plevová P, Blažek B. Intravenous immunoglobulin as prophylaxis of chemotherapy-induced oral mucositis. *J Natl Cancer Inst* 1997;89:326-7.
- [177] Nydegger U. Alte und neue Aspekte der intravenösen Immunglobulintherapie. *Schweiz Med Wochenschr* 1994;124:5-25.
- [178] Wolf HM, Eibl MM. Immunomodulatory effect of immunoglobulins. *Clin Exp Rheumatol* 1996;14 (Suppl. 15):S17-25.
- [179] Kekow J, Reinhold D, Pap T, Ansoorge S. Intravenous immunoglobulins and transforming growth factor β . *Lancet* 1998;351:184-5.
- [180] Ahmed T, Engelking C, Szalyga J, Helson L, Coombe N, Cook P, Corbi D, Puccio C, Chun H, Mittelman A. Propantheline prevention of mucositis from etoposide. *Bone Marrow Transplant* 1993;12:131-2.
- [181] Oblon DJ, Paul SR, Oblon MB, Malik S. Propantheline protects the oral mucosa after high-dose ifosfamide, carboplatin, etoposide and autologous stem cell transplantation. *Bone Marrow Transplant* 1997;20:961-3.
- [182] Sheridan WP, Morstyn G, Wolf M, Dodds A, Lusk J, Maher D, Layton JE, Green MD, Souza L, Fox RM. Granulocyte colony-stimulating factor and neutrophil recovery after high-dose chemotherapy and autologous bone marrow transplantation. *Lancet* 1989;2:891-5.
- [183] Hermann F, Schulz G, Wieser M, Kolbe K, Nicolay U, Noack M, Lindemann A, Mertelsmann R. Effect of granulocyte-macrophage colony stimulating factor on neutropenia and related morbidity induced by myelotoxic chemotherapy. *Am J Med* 1990;88:619-24.
- [184] Ho AD, Del Valle F, Haas R, Engelhard M, Hiddemann W, Ruckle H, Schlimok G, Thiel E, Andreesen R, Fiedler W. Sequential studies on the role of mitoxantrone, high-dose cytarabine, and recombinant human granulocyte-macrophage colony-stimulating factor in the treatment of refractory non-Hodgkin's lymphoma. *Semin Oncol* 1990;17 (Suppl. 10):14-19.
- [185] Grem JL, McAtee N, Murphy RF, Hamilton JM, Balis F, Steinberg S, Arbuck SG, Setser A, Jordan E, Chen A, Kohler DR, Kotite B, Allegra CJ. Phase I and pharmacokinetic study of recombinant human granulocyte-macrophage colony-stimulating factor given in combination with fluorouracil plus calcium leucovorin in metastatic gastrointestinal adenocarcinoma. *J Clin Oncol* 1994;12:560-8.
- [186] Atkinson K, Biggs JC, Downs K, Juttner C, Bradstock K, Lowenthal RM, Dale B, Szer J. GM-CSF after allogeneic bone marrow transplantation: accelerated recovery of neutrophils, monocytes and lymphocytes. *Aust NZ J Med* 1991;21:686-92.
- [187] Pettengell R, Gurney H, Radford JA, Deakin DP, James R, Wilkinson PM, Kane K, Bentley J, Crowther D. Granulocyte colony-stimulating factor to prevent dose-limiting neutropenia in non-Hodgkin's lymphoma: a randomized controlled trial. *Blood* 1992;80:1430-6.
- [188] Locatelli F, Pession A, Zecca M, Bonetti F, Prete L, Carra AM, Prete A, Montagna D, Comoli P, Taibi RM, Paolucci G. Use of recombinant human granulocyte colony-stimulating factor in children given allogeneic bone marrow transplantation for acute or chronic leukemia. *Bone Marrow Transplant* 1996;17:31-7.
- [189] Nemunaitis J, Rosenfeld CS, Ash R, Freedman MH, Deeg HJ, Appelbaum F, Singer JW, Flomenberg N, Dalton W, Elfenbein GJ. Phase III randomized, double-blind placebo-controlled trial of rhGM-CSF following allogeneic bone marrow transplantation. *Bone Marrow Transplant* 1995;15:949-54.
- [190] Katano M, Nakamura M, Matsuo T, Iyama A, Hisatsugu T. Effect of granulocyte colony-stimulating factor (G-CSF) on chemotherapy-induced oral mucositis. *Surg Today* 1995;25:202-6.
- [191] Kannan V, Bapsy PP, Anantha N, Doval DC, Vaithianathan H, Banumathy G, Reddy KB, Kumaraswamy SV, Shenoy AM. Efficacy and safety of granulocyte macrophage-colony stimulating factor (GM-CSF) on the frequency and severity of radiation mucositis in patients with head and neck carcinoma. *Int J Radiat Oncol Biol Phys* 1997;37:1005-10.
- [192] Rosso M, Blasi G, Gherlone E, Rosso R. Effect of granulocyte-macrophage colony-stimulating factor on prevention of mucositis in head and neck cancer patients treated with chemoradiotherapy. *J Chemother* 1997;9:382-5.
- [193] Keith JC, Jr, Albert L, Sonis ST, Pfeiffer CJ, Schaub RG. IL-11, a pleiotropic cytokine: exciting new effects of IL-11 on gastrointestinal mucosal biology. *Stem Cells* 1994;12 (Suppl. 1):79-89.
- [194] Sonis S, Muska A, O'Brien J, Van Vugt A, Langer-Safer P, Keith J. Alteration in the frequency, severity and duration of chemotherapy-induced mucositis in hamsters by interleukin-11. *Eur J Cancer B Oral Oncol* 1995;31B:261-6.
- [195] Sonis ST, Van Vugt AG, McDonald J, Dotoli E, Schwertschlag U, Szkut P, Keith J. Mitigating effects of interleukin 11 on consecutive courses of 5-fluorouracil-induced ulcerative mucositis in hamsters. *Cytokine* 1997;9:605-12.
- [196] Rand KH, Kramer B, Johnson AC. Cancer chemotherapy associated symptomatic stomatitis: role of HSV (herpes simplex virus). *Cancer* 1982;50:1262-5.
- [197] Gluckman E, Lotsberg J, Devergie A, Zhao XM, Melo R, Gomez-Morales M, Mazon MC, Perol Y. Oral acyclovir prophylactic treatment of herpes simplex infection after bone marrow transplantation. *J Antimicrob Chemother* 1983;12 (Suppl. B):161-7.
- [198] Greenberg MS, Cohen SG, Boosz B, Griedman H. Oral herpes simplex infection in patients with leukemia. *J Am Dent Assoc* 1987;114:483-6.
- [199] Redding SW. Role of herpes simplex virus reactivation in chemotherapy-induced oral mucositis. *NCI Monogr* 1990;9:103-5.
- [200] Redding SW, Montgomery MT. Acyclovir prophylaxis for oral herpes simplex virus infection in patients with bone marrow transplants. *Oral Surg* 1989;67:680-3.
- [201] Woo SB, Sonis ST, Sonis AL. The role of herpes simplex virus in the development of oral mucositis in bone marrow transplant recipients. *Cancer* 1990;66:2375-9.
- [202] Epstein JB, Ransier A, Sherlock CH, Spinelli JJ, Reece D. Acyclovir prophylaxis of oral herpes virus during bone marrow transplantation. *Eur J Cancer B Oral Oncol* 1996;32B:158-62.
- [203] Redding SW, Montgomery MT. Acyclovir prophylaxis for oral herpes simplex virus infection in patients with bone marrow transplants. *Oral Surg Oral Med Oral Pathol* 1989;67:680-3.
- [204] Bubley GJ, Chapman B, Chapman SK, Crumacker CS, Schnipper LE. Effect of Acyclovir on radiation- and chemotherapy-induced mouth lesions. *Antimicrob Agents Chemother* 1989;33:862-5.

- [205] Bergmann OJ, Mogensen SC, Ellermann-Eriksen S, Ellegaard J. Acyclovir prophylaxis and fever during remission-induction therapy of patients with acute myeloid leukemia: a randomized, double blind, placebo-controlled trial. *J Clin Oncol* 1997;15: 2269-74.
- [206] Newsholme EA, Newsholme P, Curi R, Challoner E, Ardawi MSM. A role for muscle in the immune system and its importance in surgery, trauma, sepsis and burns. *Nutrition* 1988; 4:261-8.
- [207] Fürst P. Intracellular muscle free amino-acids; their measurement and function. *Proc Nutr Soc* 1983;42:451-62.
- [208] Fürst P, Stehle P. The potential use of parenteral dipeptides in clinical nutrition. *Nutr Clin Pract* 1993;8:106-14.
- [209] van der Hulst RRWJ, van Kreel BK, von Meyenfeldt MF, Brummer RJM, Arends JW, Deutz NEP, Soeters PB. Glutamine and the preservation of gut integrity. *Lancet* 1993; 341:1363-5.
- [210] Higashiguchi T, Hasselgren PO, Wagner K, Fischer JE. Effect of glutamine on protein synthesis in isolated intestinal epithelial cells. *J Parenter Enteral Nutr* 1993;17:307-14.
- [211] Fox AD, Kripke SA, de Paula JA, Berman JM, Settle RG, Rombeau JL. Effect of a glutamine-supplemented enteral diet on methotrexate-induced enterocolitis. *J Parent Ent Nutr* 1988;12:325-31.
- [212] Skubitz KM, Anderson PM. Oral glutamine to prevent chemotherapy-induced stomatitis: a pilot study. *J Lab Clin Med* 1996;127:223-8.
- [213] Anderson PM, Schroeder G, Skubitz KM. Oral glutamine reduces the duration and severity of stomatitis after cytotoxic cancer chemotherapy. *Cancer* 1998;83:1433-9.
- [214] van Zaanen HCT, van der Lelie H, Timmer JG, Fürst P, Sauerwein H. Parenteral glutamine dipeptide supplementation does not ameliorate chemotherapy-induced toxicity. *Cancer* 1994;74:2879-84.
- [215] Jebb SA, Osborne RJ, Maughan TS, Mohideen N, Mack P, Mort D, Shelley MD, Elia M. 5-fluorouracil and folinic acid-induced mucositis: no effect of oral glutamine supplementation. *Br J Cancer* 1994;70:732-5.
- [216] Lissoni P, Tancini G, Barni S, Paolorossi F, Ardizzio A, Conti A, Maestroni G. Treatment of cancer chemotherapy-induced toxicity with the pineal hormone melatonin. *Support Care Cancer* 1997;5:126-9.
- [217] Wolfrom C, Hartmann R, Fengler R, Bruhmüller S, Ingwersen A, Henze G. Randomized comparison of 36-hour intermediate dose versus 4-hour high-dose methotrexate infusions for remission induction in relapsed childhood acute lymphoblastic leukemia. *J Clin Oncol* 1993;11:827-33.
- [218] Lévi F. Chronopharmacology of anticancer agents. In: Redfern PH, Lemmer B, editors. *Handbook of Experimental Pharmacology: Physiology and Pharmacology of Biological Rhythms: Cancer Chemotherapy*. Berlin: Springer-Verlag, 1997. p. 299-331.
- [219] Smaaland R, Laerum OD, Lote K, Sletvold O, Sothorn RB, Bjerknes R. DNA synthesis in human bone marrow is circadian stage dependent. *Blood* 1991;77:2603-11.
- [220] Smaaland R, Abrahamsen JF, Svoldal AM, Lote K, Ueland PM. DNA cell cycle distribution and glutathione (GSH) content according to circadian stage in bone marrow of cancer patients. *Br J Cancer* 1992;66:39-45.
- [221] Lévi F, Zidani R, Misset JL. Randomised multicentre trial of chronotherapy with oxaliplatin, fluorouracil, and folinic acid in metastatic colorectal cancer. *Lancet* 1997;350:681-6.
- [222] Jansma J, Vissink A, Bouma J, Vermey A, Panders AK, 's-Gravenmade EJ. A survey of prevention and treatment regimens for oral sequelae resulting from head and neck radiotherapy used in Dutch radiotherapy institutes. *Int J Radiat Oncol Biol Phys* 1992;24:359-67.
- [223] Mueller BA, Millheim ET, Farrington EA, Brusko C, Wiser TH. Mucositis management practices for hospitalized patients: national survey results. *J Pain Symptom Manage* 1995;10:510-20.
- [224] Zimmermann JS, Wilhelm R, Niehoff P, Schneider R, Kovacs G, Kimmig B. Prophylaxe und Therapie akuter Strahlenfolgen an Haut und Schleimhaut. Teil I. Ergebnisse einer bundesweiten Erhebung. *Strahlenther Onkol* 1998;174:142-8.
- [225] Jacobson JM, Greenspan JS, Spritzler J, Ketter N, Fahey JL, Jackson JB, Fox L, Chernoff M, Wu AW, MacPhail LA, Vasquez GJ, Wohl DA. Thalidomide for the treatment of oral aphthous ulcers in patients with human immunodeficiency virus infection. *N Engl J Med* 1997; 336:1487-93.